

"Bacterial" Genome structures

4BIM - INSA 2006

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18 juin 2006

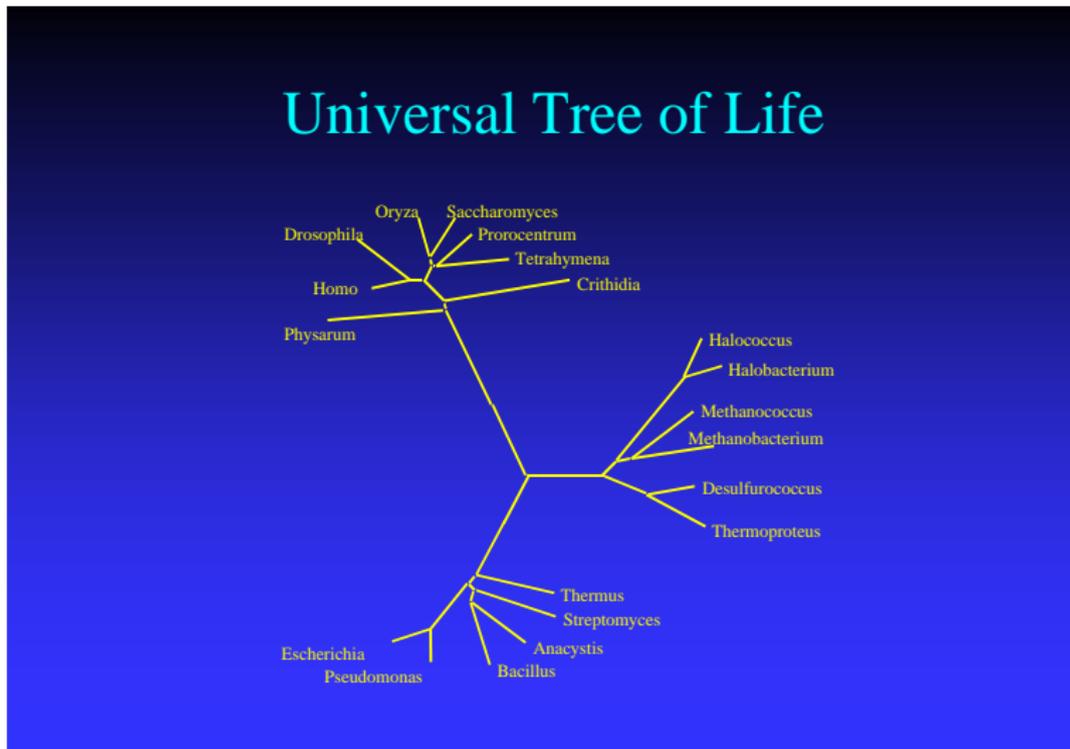
Outline

- 1 Introduction
- 2 Genome size
- 3 Topology & #
- 4 G+C content
- 5 Replichores
- 6 Gene orientation biases
- 7 Chirochores
- 8 X-rated structure

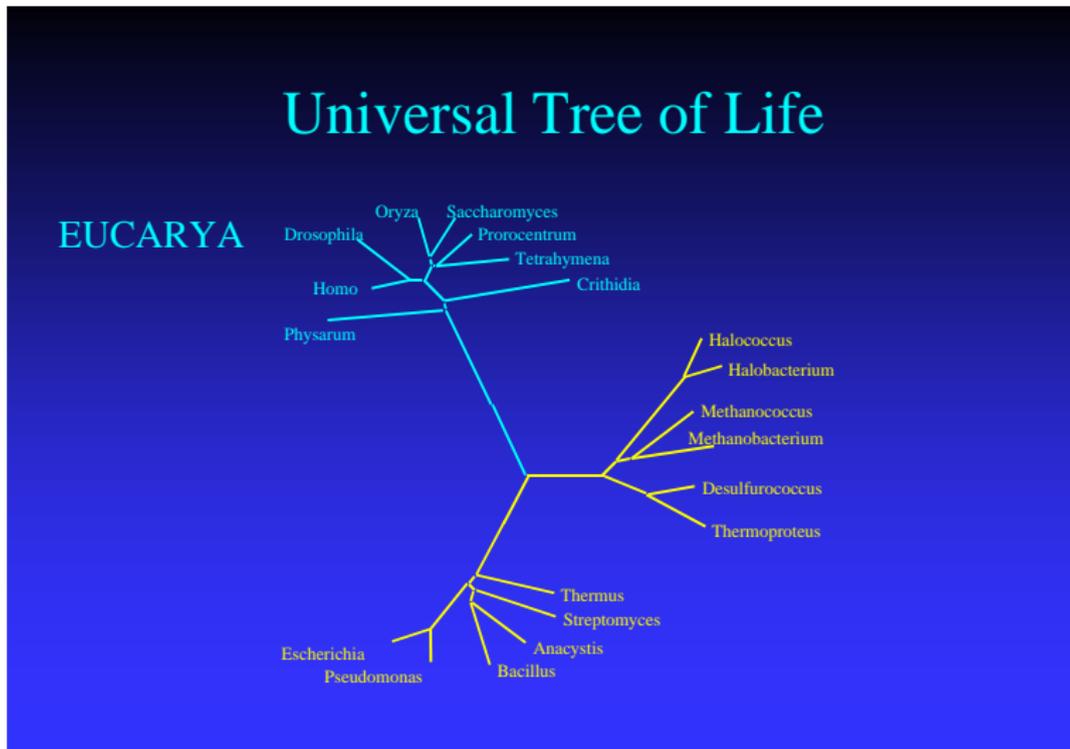
Introduction

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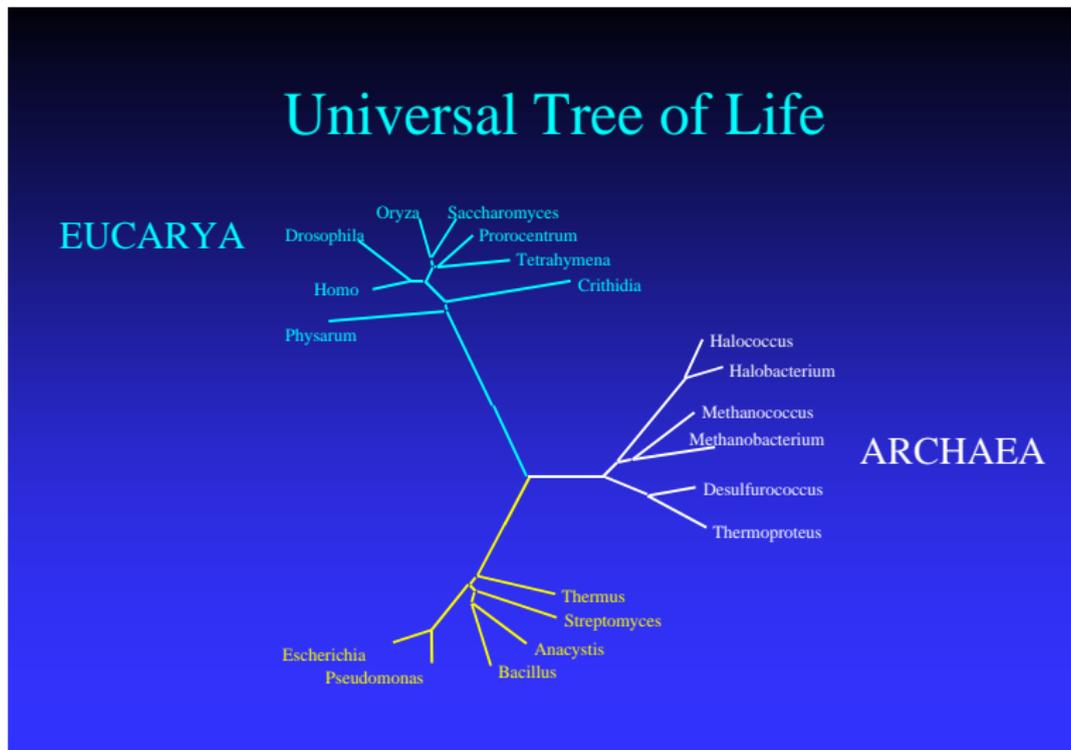
The three kingdoms



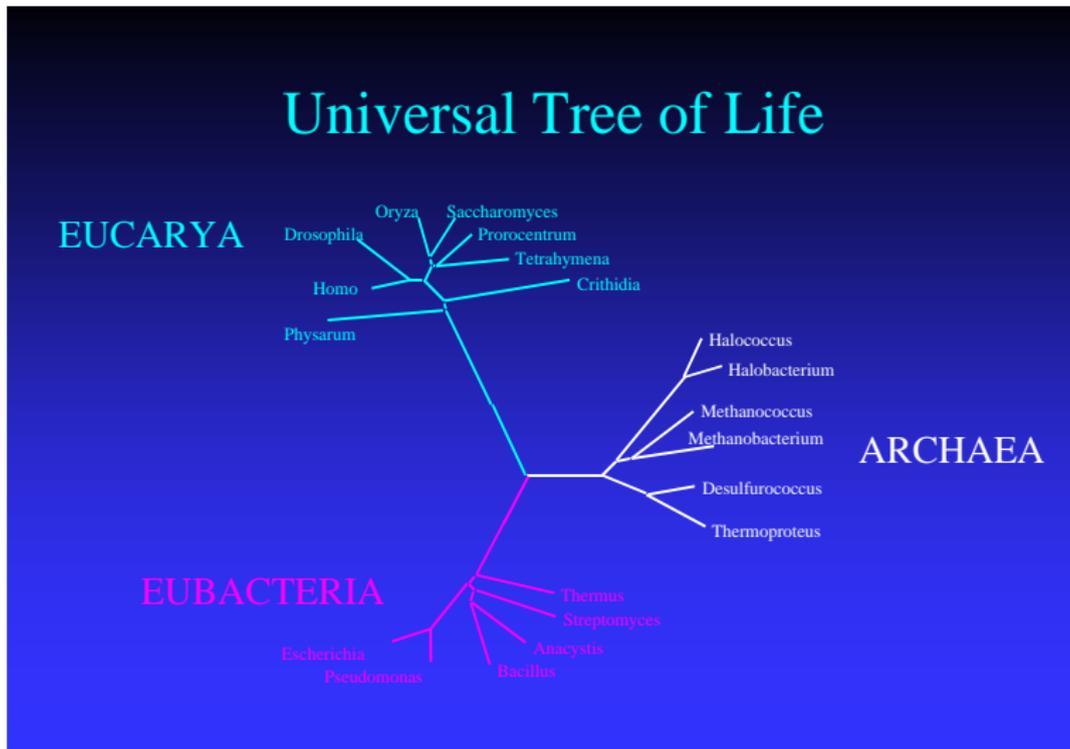
The three kingdoms : eucarya



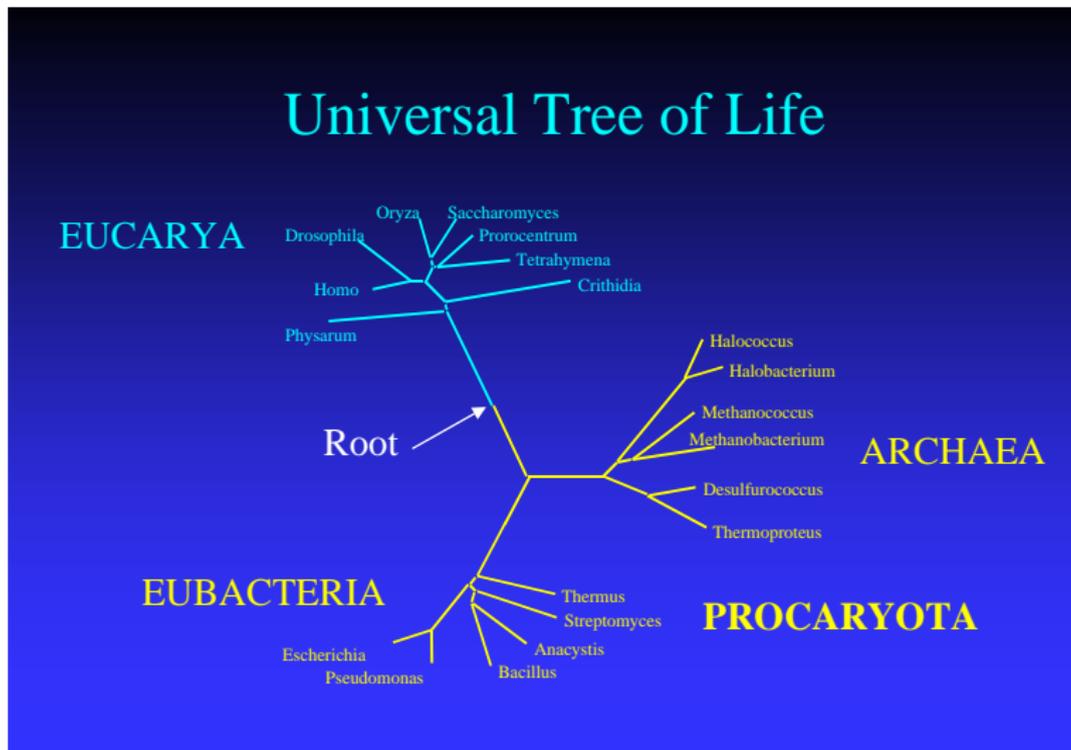
The three kingdoms : archaea



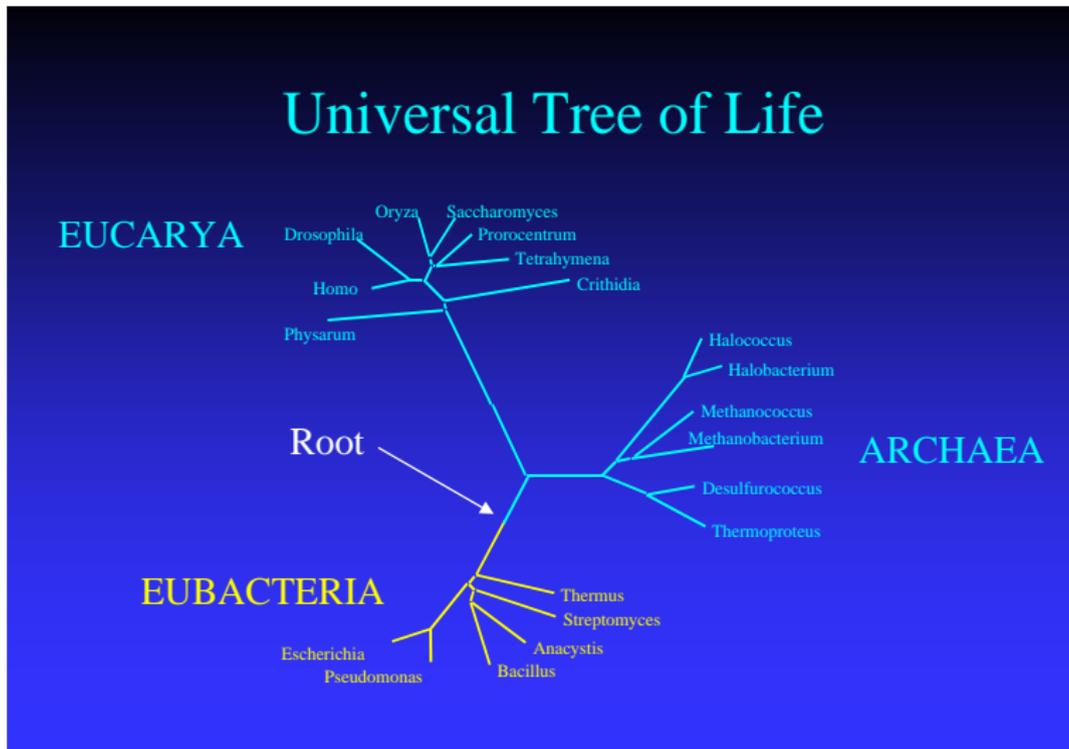
The three kingdoms : eubacteria



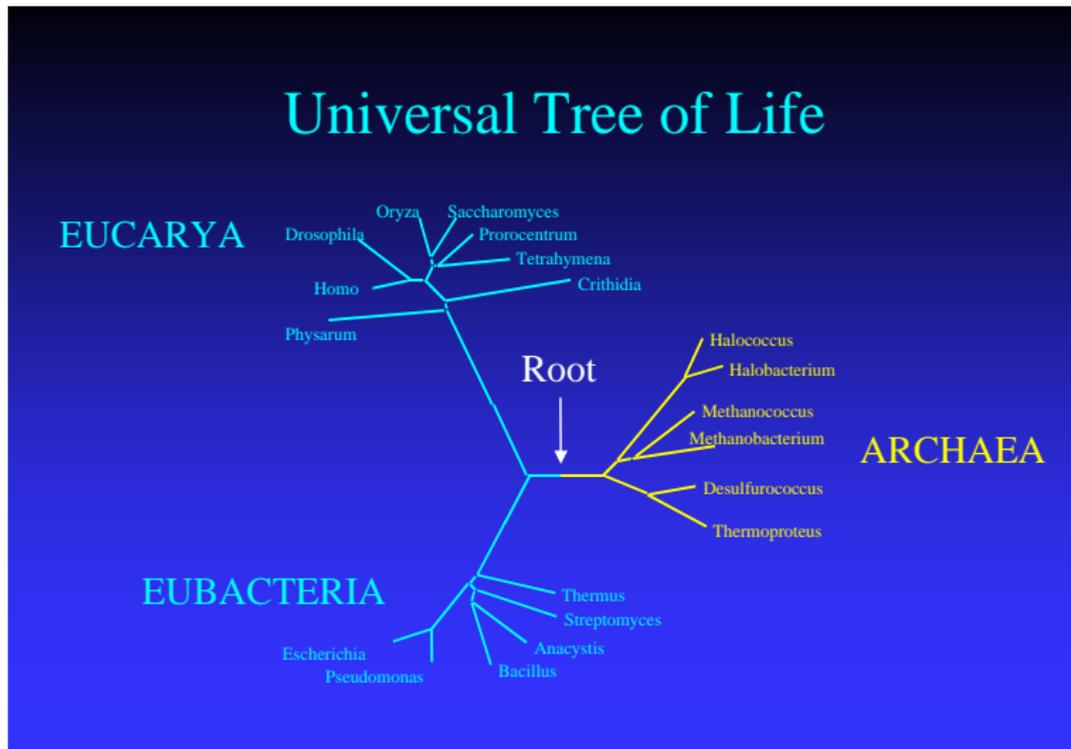
The three kingdoms : root 1



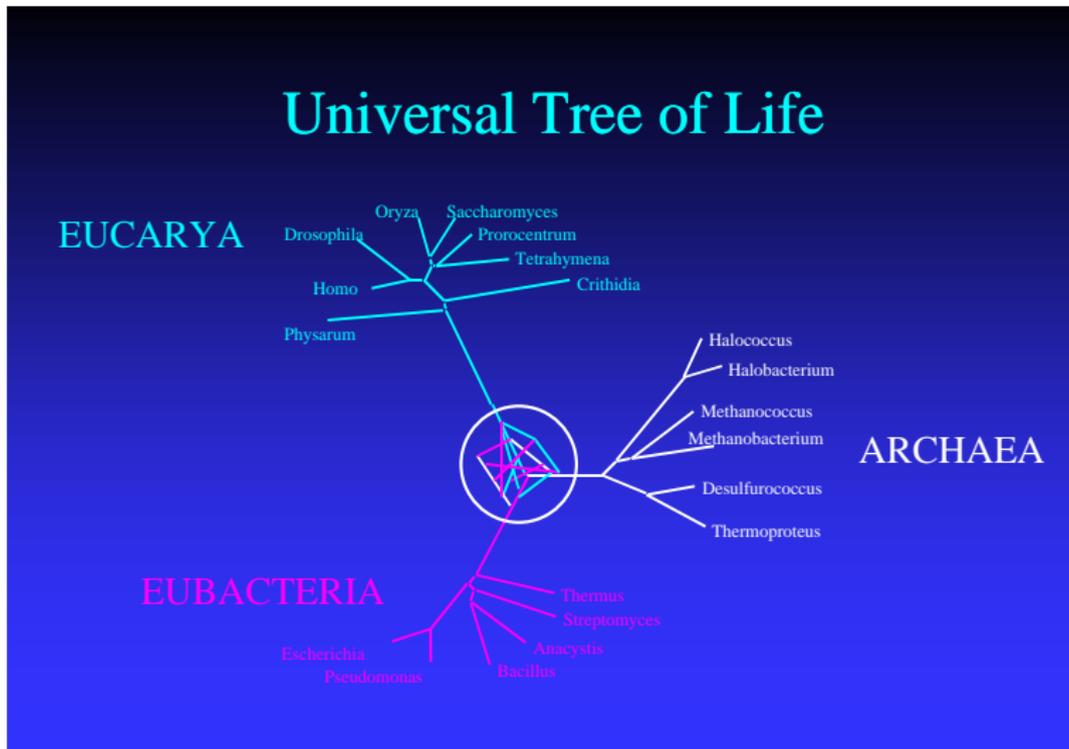
The three kingdoms : root 2



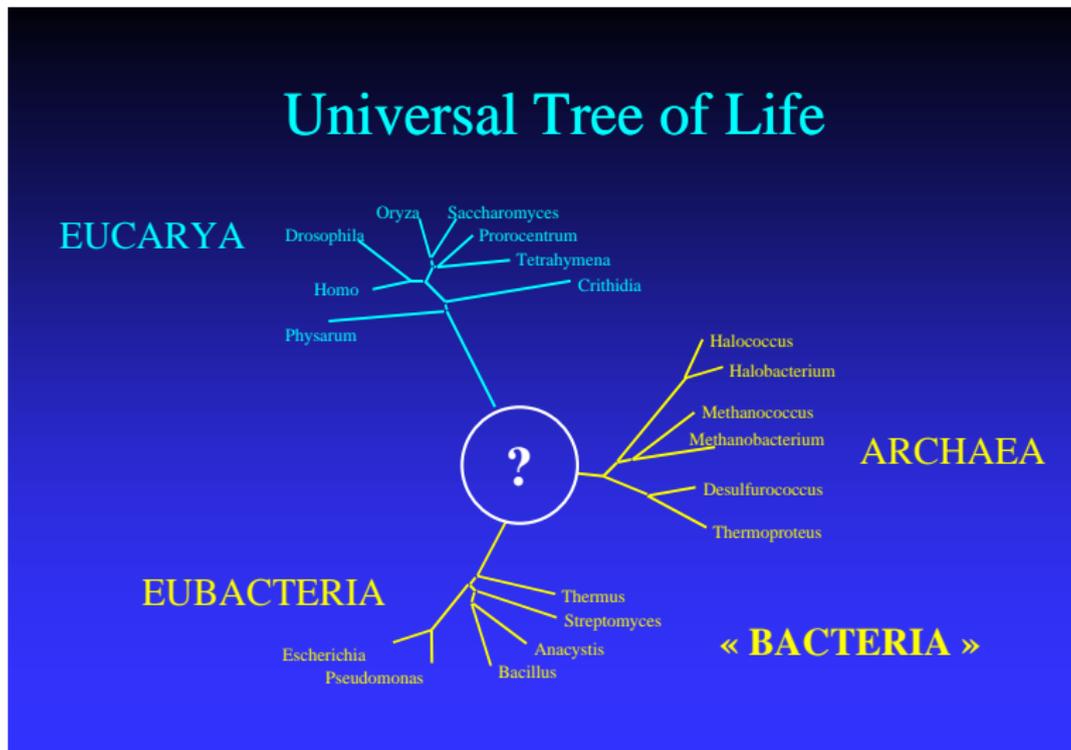
The three kingdoms : root 3



The three kingdoms : no root



The three kingdoms : "bacteria"



Organelles : chloroplasts & mitochondria

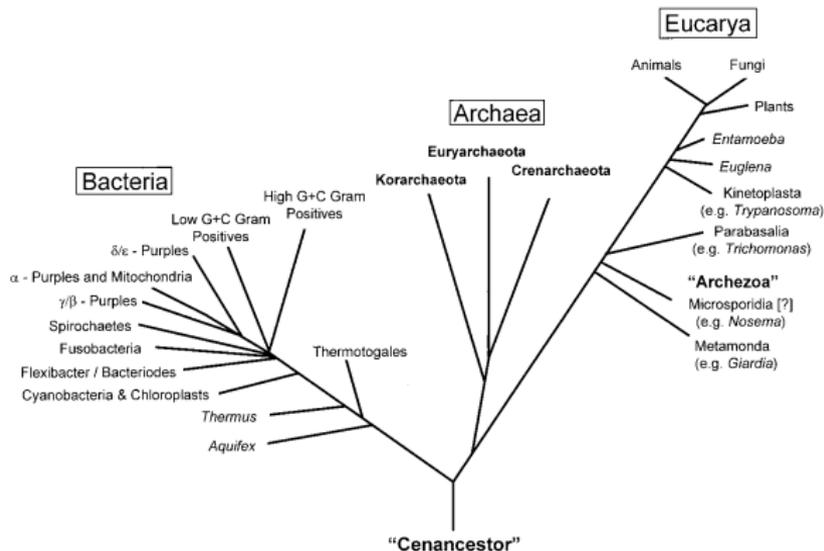
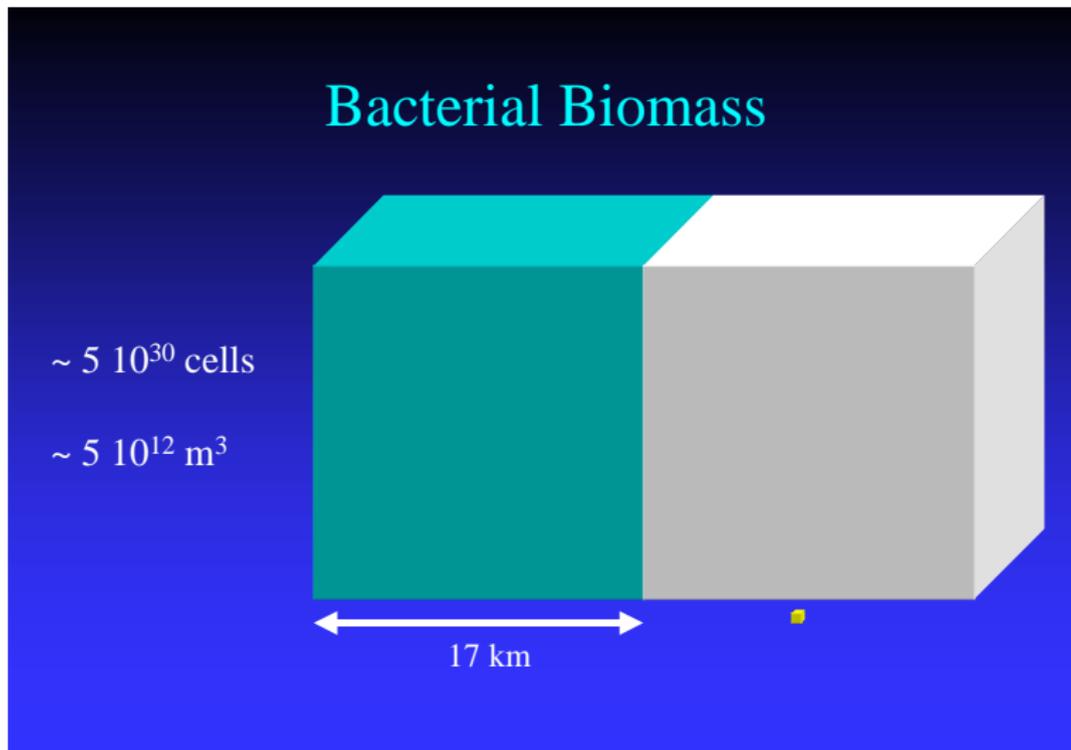
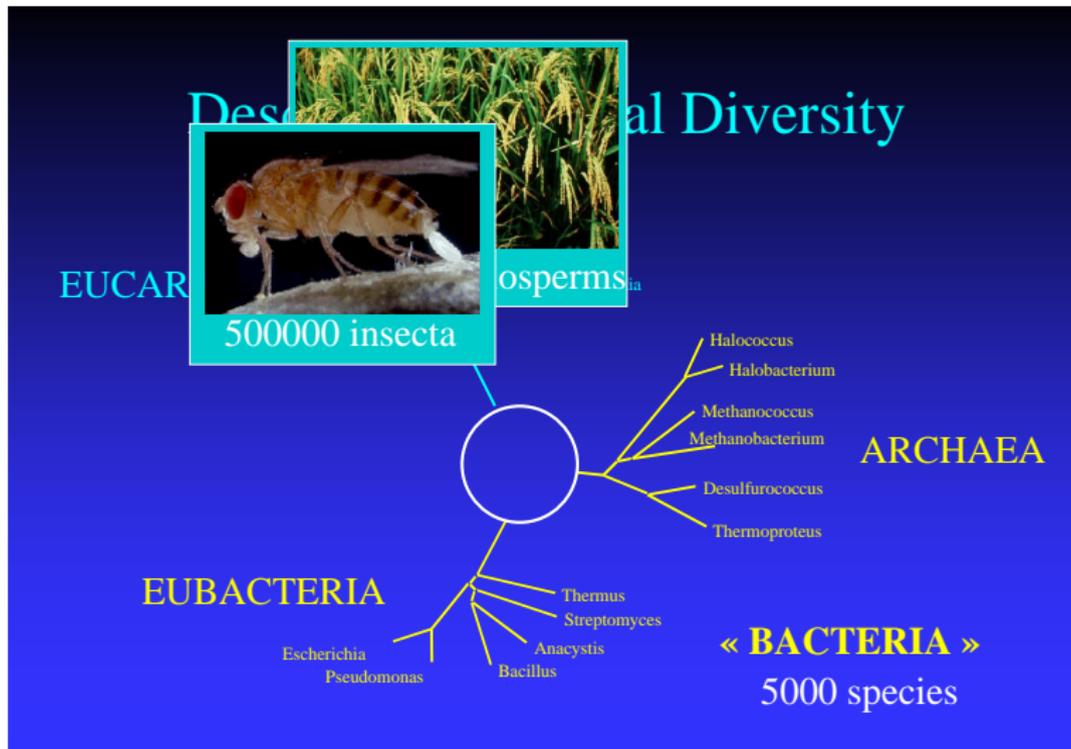


FIG. 1. Schematic drawing of a universal rRNA tree showing the relative positions of evolutionary pivotal groups in the domains *Bacteria*, *Archaea*, and *Eucarya*. The location of the root (the cenacestor) corresponds to that proposed by reciprocally rooted gene phylogenies (43, 133, 164). The question mark beside the Archezoa group Microsporidia denotes recent suggestions that it might branch higher in the eukaryotic portion of the tree. (Branch lengths have no meaning in this tree.)

Half of biomass on earth



Very few species



First species classification

An old trend

Aristotle (-384,-322)

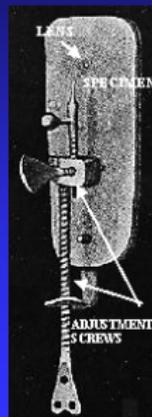


508 Eucaryota
0 Bacteria

Small bacteria

Little Bacteria

Antony van Leeuwenhoek (1632-1723)



$$0.1 \text{ mm} = 100 \mu\text{m}$$



Thickness $\approx 100 \mu\text{m}$.

1 M€ $\approx 1 \text{ m}$

1 G€ $\approx 1 \text{ km}$

Bacterial cell size is in μm



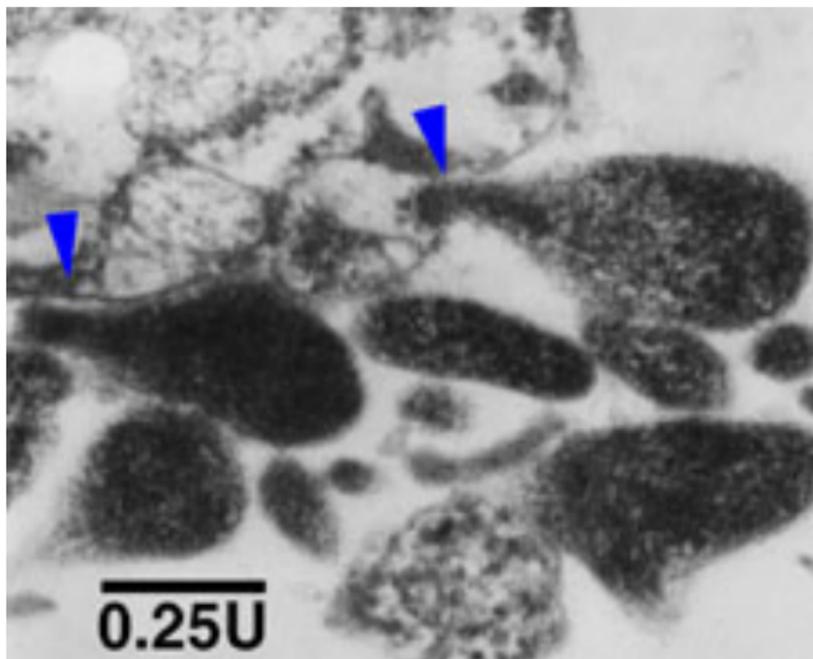
Demo

A giant : *Epulopiscium fishelsoni* bar is 50 μm



Bresler, V. *et al* (1998) *J. Bact.*, **180** :5601-5611.

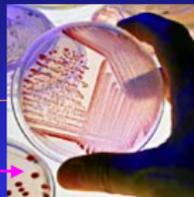
Mycoplasma genitalium bar is 0.25 μm



1 colony $\approx 10^6$ cells

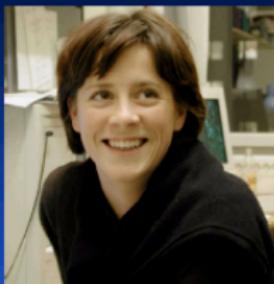
Little Bacteria

1 colony 10^6 cells



Few morphological traits

Ugly Little Bacteria

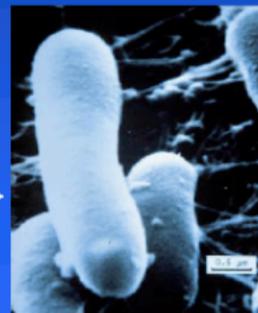


10^7 years



E. coli

10^9 years



B. subtilis

Bacterial classification

Classification of Bacteria

- Was based mainly on physiology and growth conditions



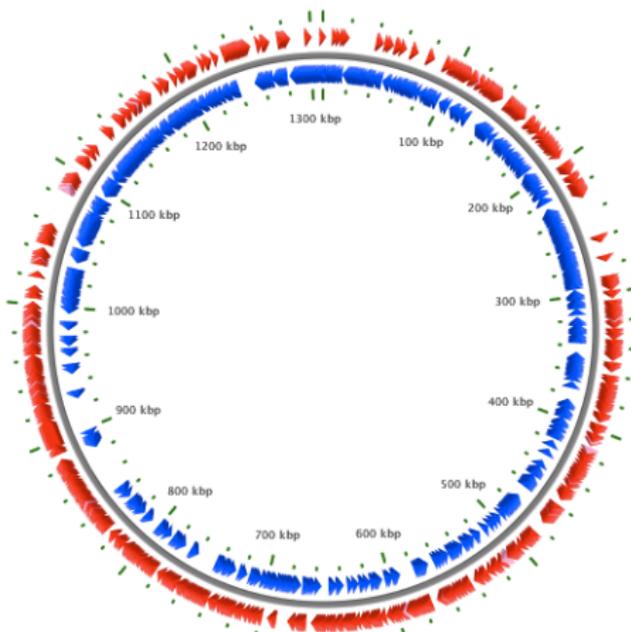
E. coli



Most bacteria defy cultivation...

Bacterial classification : "*Candidatus Pelagibacter ubique*"

Candidatus Pelagibacter ubique HTCC1062, complete genome



Bacterial classification : *Candidatus*

Candidatus examples :

- "*Candidatus* Arsenophonus triatominarum"
- "*Candidatus* Arthromitus"
- "*Candidatus* Blochmannia"
- "*Candidatus* Blochmannia floridanus"
- "*Candidatus* Blochmannia herculeanus"
- "*Candidatus* Burkholderia kirkii"
- "*Candidatus* Glomeribacter gigasporarum"
- "*Candidatus* Xiphinematobacter brevicollis"

Genome size

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bp : base pair

Common multiples are :

- 1 kb = 10^3 bp
- 1 Mb = 10^6 bp
- 1 Gb = 10^9 bp

Bacterial genomes are typically expressed in Mb

Length conversion

Dickerson *et al* (1982) *Science*, **216** :475-485.

1 bp \approx 0.33 nm

- 1 kb \approx 0.33 μ m
- 1 Mb \approx 0.33 mm
- 1 Gb \approx 0.33 m

Bacterial genomes are typically in the **mm** range.

Mass conversion ($1 \text{ pg} = 10^{-12} \text{ g}$)

Doležel *et al* (2003) *Cytometry*, **51A** :127-128.

Number of base pairs = mass in pg $\times 0.978 \times 10^9$

- $1 \text{ kb} \approx 10^{-6} \text{ pg}$
- $1 \text{ Mb} \approx 10^{-3} \text{ pg}$
- $1 \text{ Gb} \approx 1 \text{ pg}$

Bacterial genomes are typically in the 10^{-3} pg range (femtogram).

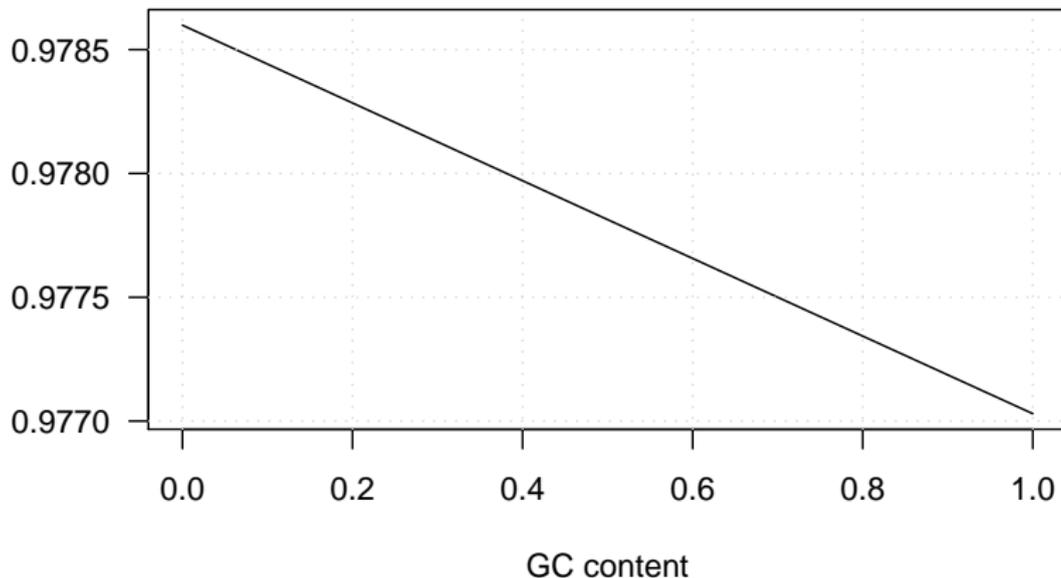
Mass conversion constant and G+C content

Base	Nucleotide	Chemical formula
A	2'-deoxyadenosine 5'-monophosphate	$C_{10}H_{14}N_5O_6P$
T	2'-deoxythymidine 5'-monophosphate	$C_{10}H_{15}N_2O_8P$
G	2'-deoxyguanosine 5'-monophosphate	$C_{10}H_{14}N_5O_7P$
C	2'-deoxycytidine 5'-monophosphate	$C_9H_{14}N_3O_7P$

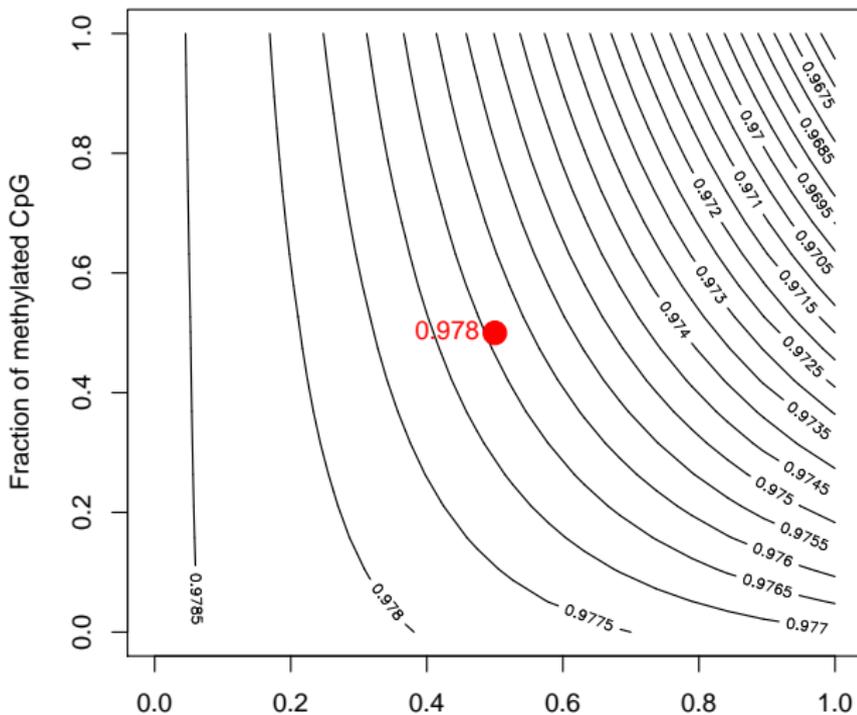
Tab.: Chemical formula of the four nucleotides in DNA.

Mass conversion constant and G+C content

Evolution of the conversion constant with GC content



Evolution of the conversion constant with GC content and the fraction of methylated CpG



The big picture

Virus, organelles

Tiny genomes (kb)

High gene density

"Bacteria"

Small genomes (Mb)

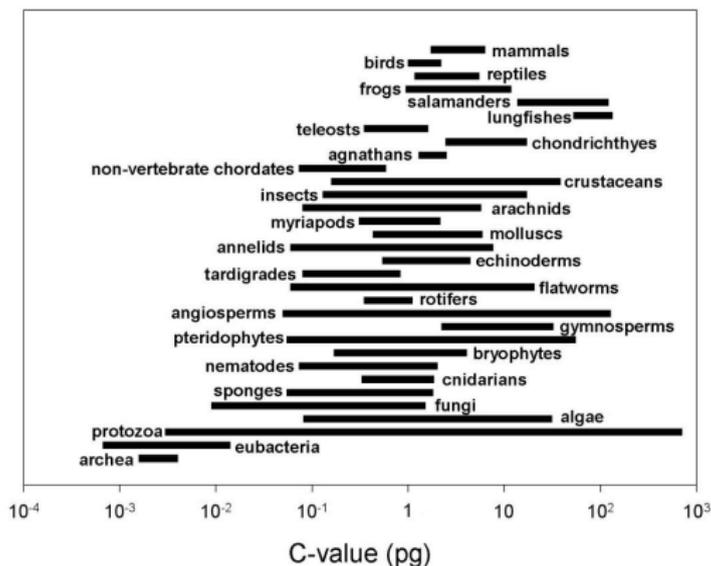
High gene density

Eucarya

Large genomes (Gb)

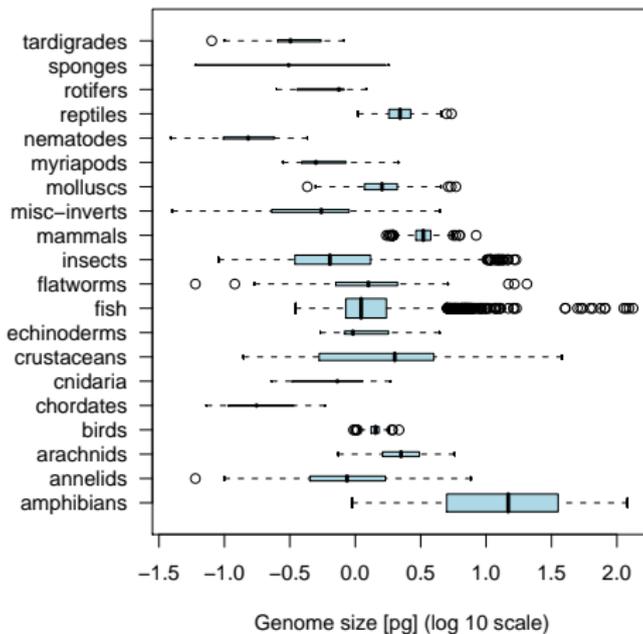
Low gene density

C value paradox

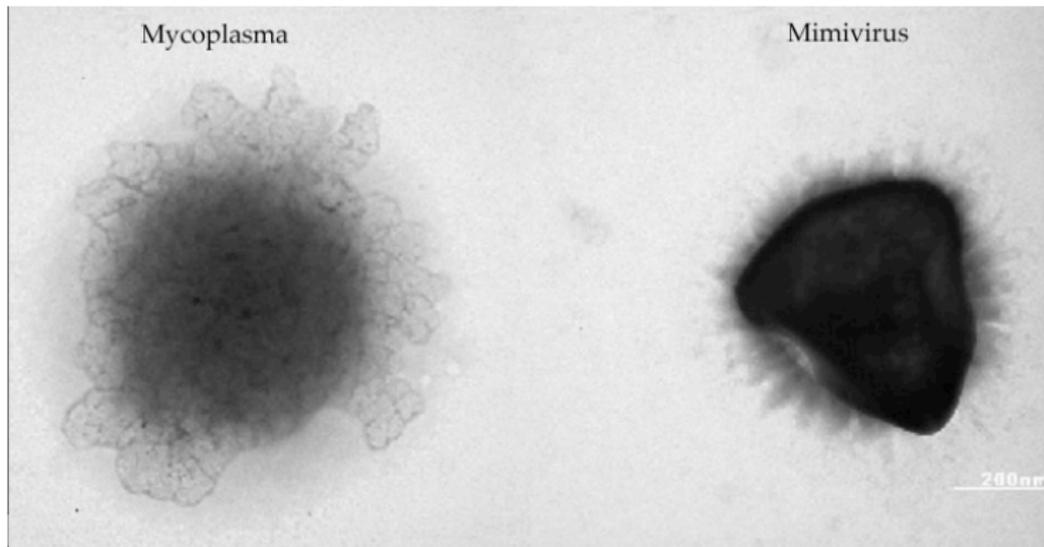


Gregory, T.R. (2004) *Paleobiology*, **30** :179-202.

C value paradox

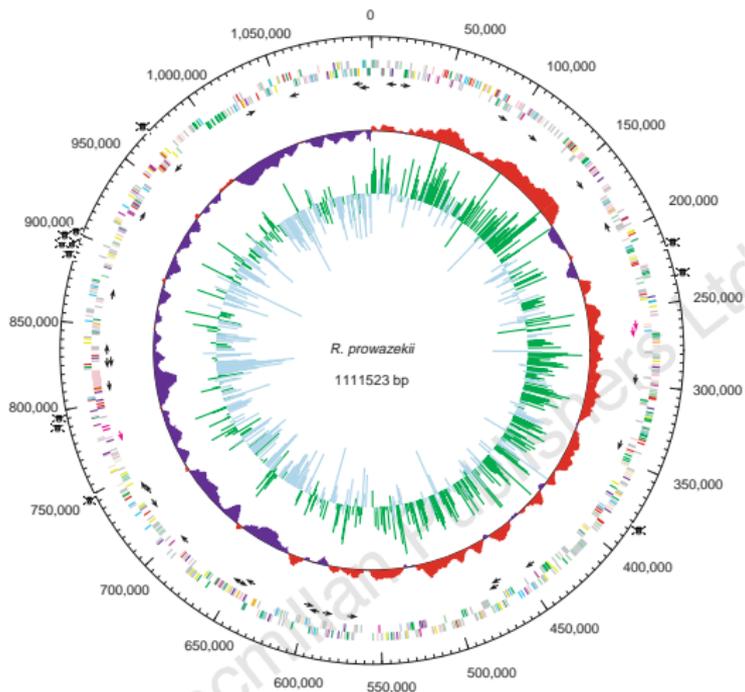
Gregory, T.R. (2005) *Animal Genome Size Database*

Giant virus : mimivirus 1.2 Mb



Electronic microscopy of a "bacteria" on the left (*Ureaplasma urealyticum (parvum)*) with a genome size of 0.751 Mb and mimivirus on the right with a genome size of 1.181 Mb. Credit : the Mimivirus picture gallery from <http://giantvirus.org/>. Copyright : Prof. Didier Raoult, Rickettsia Laboratory, La Timone, Marseille, France.

Pseudogenes in *Rickettsia prowazekii*



Pseudogenes in *Mycobacterium leprae*

Massive gene decay in the leprosy bacillus

S. T. Cole*, K. Eiglmeier*, J. Parkhill†, K. D. James†, N. R. Thomson†, P. R. Wheeler‡, N. Honoré*, T. Garnier*, C. Churcher†, D. Harris†, K. Mungall†, D. Basham†, D. Brown†, T. Chillingworth†, R. Connor†, R. M. Davies†, K. Devlin†, S. Duthoy*, T. Feltwell†, A. Fraser†, N. Hamlin†, S. Holroyd†, T. Hornsby†, K. Jagels†, C. Lacroix†, J. Maclean†, S. Moule†, L. Murphy†, K. Oliver†, M. A. Quail†, M.-A. Rajandream†, K. M. Rutherford†, S. Rutter†, K. Seeger†, S. Simon*, M. Simmonds†, J. Skelton†, R. Squares†, S. Squares†, K. Stevens†, K. Taylor†, S. Whitehead†, J. R. Woodward† & B. G. Barrell†

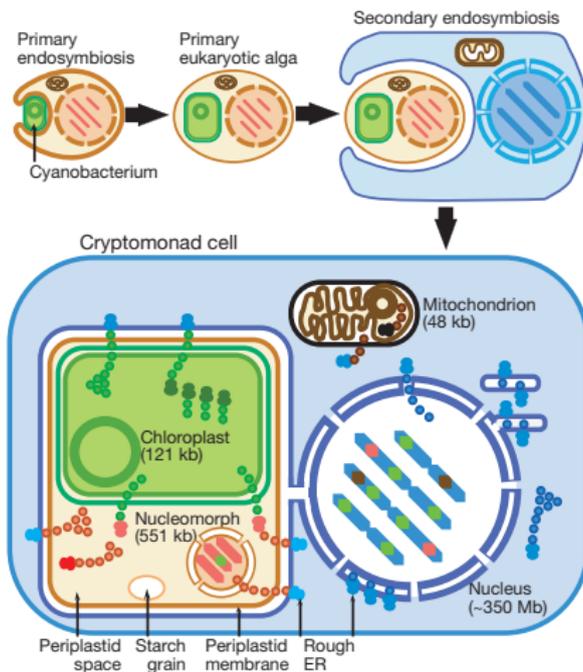
*Unité de Génétique Moléculaire Bactérienne, Institut Pasteur, 28 rue du Docteur Roux, 75724 Paris Cedex 15, France

†Sanger Centre, Wellcome Trust Genome Campus, Hinxton, CB10 1SA, UK

‡Veterinary Laboratories Agency, Weybridge, Woodham Lane, New Haw, Addlestone, Surrey KT15 3NB, UK

Leprosy, a chronic human neurological disease, results from infection with the obligate intracellular pathogen *Mycobacterium leprae*, a close relative of the tubercle bacillus. *Mycobacterium leprae* has the longest doubling time of all known bacteria and has thwarted every effort at culture in the laboratory. Comparing the 3.27-megabase (Mb) genome sequence of an armadillo-derived Indian isolate of the leprosy bacillus with that of *Mycobacterium tuberculosis* (4.41 Mb) provides clear explanations for these properties and reveals an extreme case of reductive evolution. Less than half of the genome contains functional genes but pseudogenes, with intact counterparts in *M. tuberculosis*, abound. Genome downsizing and the current mosaic arrangement appear to have resulted from extensive recombination events between dispersed repetitive sequences. Gene deletion and decay have eliminated many important metabolic activities including siderophore production, part of the oxidative and most of the microaerophilic and anaerobic respiratory chains, and numerous catabolic systems and their regulatory circuits.

Tiny eucaryal genome : *Guillardia theta* is only 551 kb



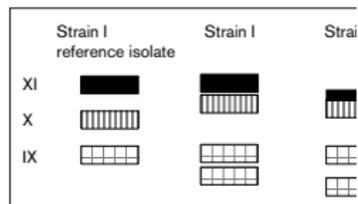
Tiny eucaryal genome : *Encephalitozoon cuniculi* is only 2.9 Mb

Towards the minimal eukaryotic parasitic genome

Christian P Vivarès* and Guy Méténier

Microsporidia are well-known to infect immunocompromised patients and are also responsible for clinical syndromes in immunocompetent individuals. In recent years, evidence has been obtained in support of a very close relationship between Microsporidia and Fungi. In some species, the compaction of the genome and genes is remarkable. Thus, a systematic sequencing project has been initiated for the 2.9 Mbp genome of *Encephalitozoon cuniculi*, which will be useful for future comparative genomic studies.

Figure 1



Katinka, M.D. *et al* (2001) *Nature*, **414** :450-453.

Overlap of free living forms

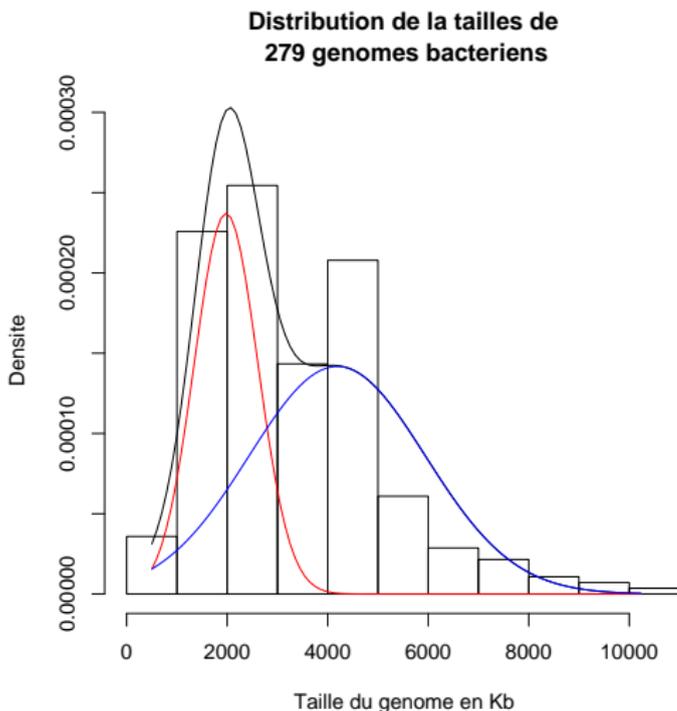
- Eucarya *Saccharomyces cerevisiae* is 12 Mb
- Bacteria *Sorangium cellulosum* is 13 Mb

What is the distribution of bacterial genome size ?

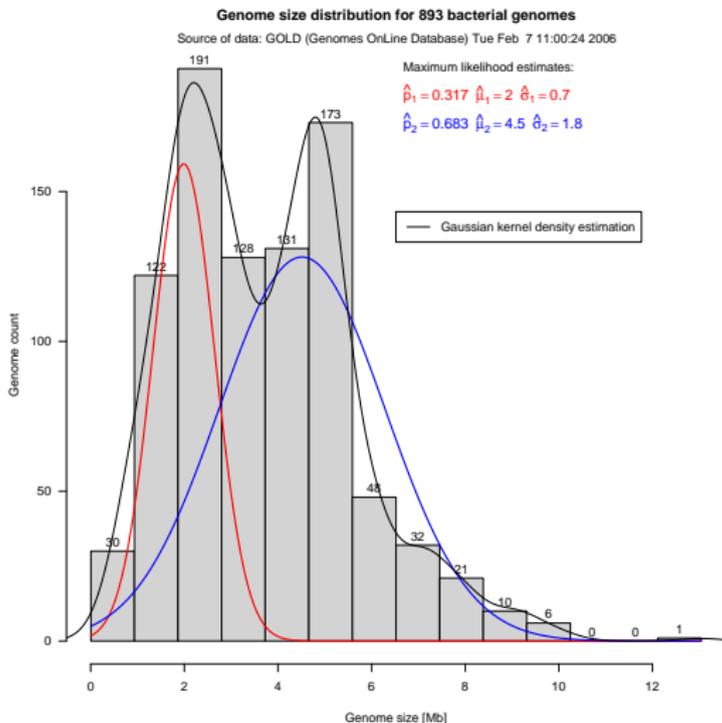
Study this yourself :

<http://pbil.univ-lyon1.fr/R/fichestd/tdr222.pdf>

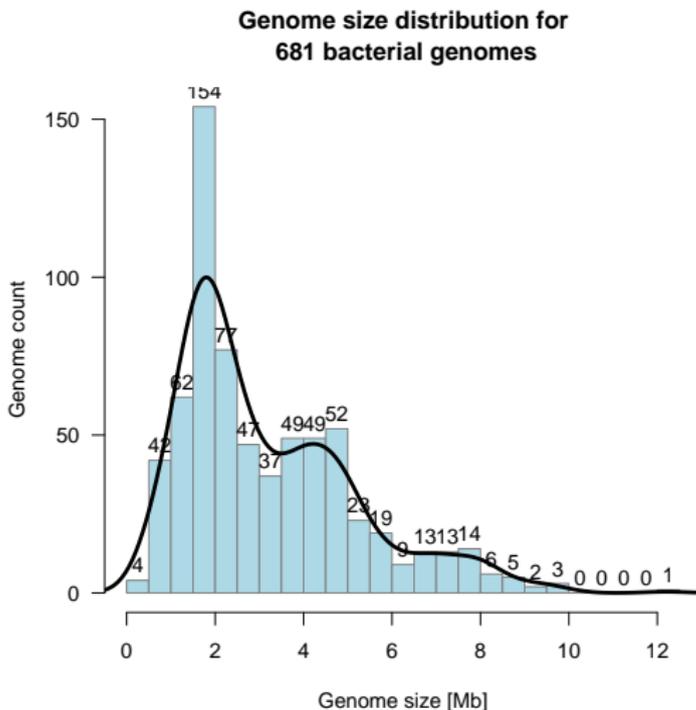
Genome size for 279 bacteria (GOLD 2002)



Genome size for 893 bacteria (GOLD 2006)



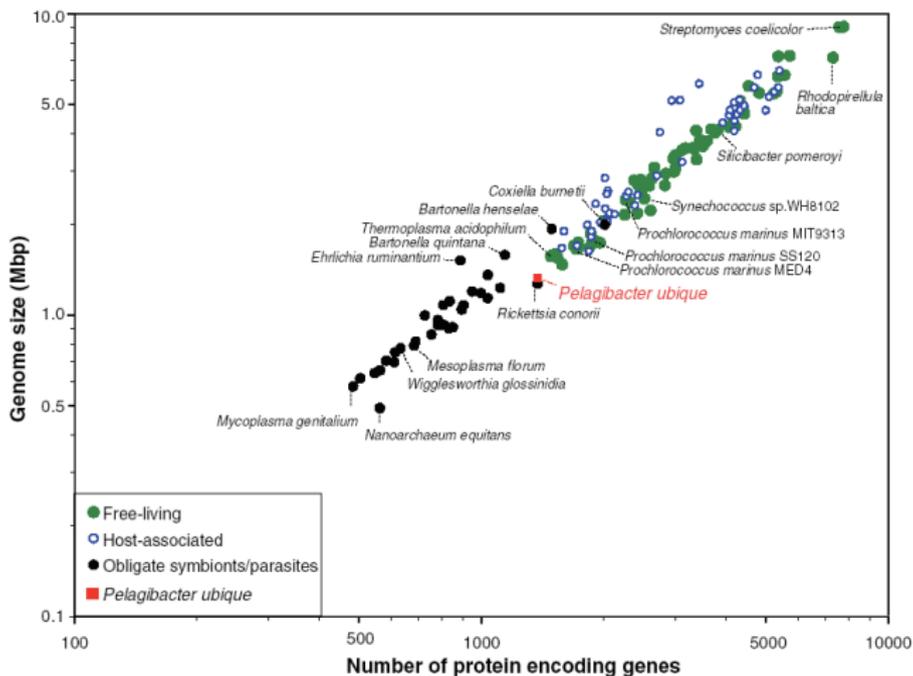
Genome size for 681 bacteria (PFGE data)



Genome size summary

- Range : from 0.45 Mb (*Buchnera*) to 13.0 Mb (*Sorangium cellulosum*).
- Three modes at 2 Mb, 4.5 Mb, and 8 Mb, respectively.

Generalists versus specialists



Giovannoni, S.J. *et al* (2005) *Science*, **309** :1242-1245.

Genome size & repeat density



Genetica **115**: 1–12, 2002.
© 2002 Kluwer Academic Publishers. Printed in the Netherlands.

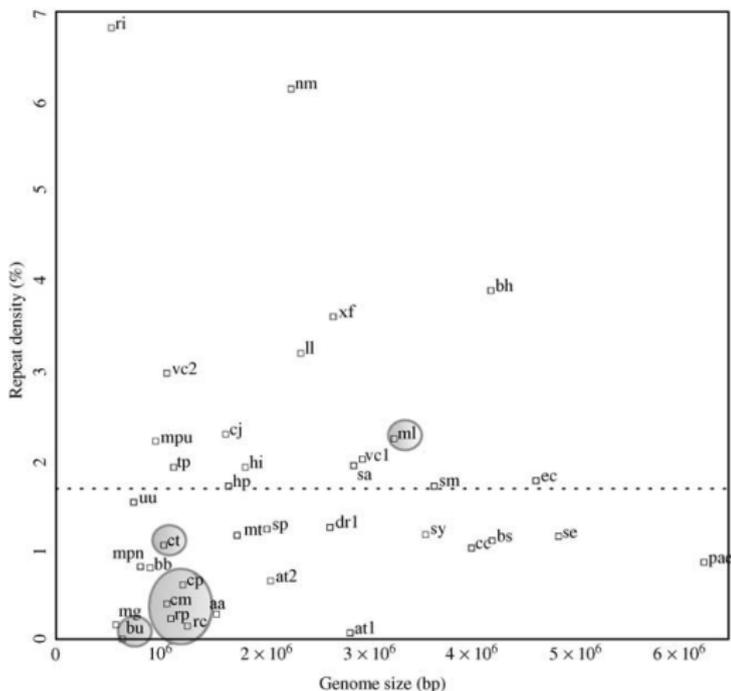
1

Genome deterioration: loss of repeated sequences and accumulation of junk DNA

A. Carolin Frank, Haleh Amiri & Siv G.E. Andersson*

*Department of Molecular Evolution, University of Uppsala, Uppsala, S-751 36 Sweden; *Author for correspondence (Phone: +46-18-4714379; Fax: +46-18-471 64 04; E-mail: Siv.Andersson@ebc.uu.se)*

Genome size & repeat density



Genome size & repeat density

8

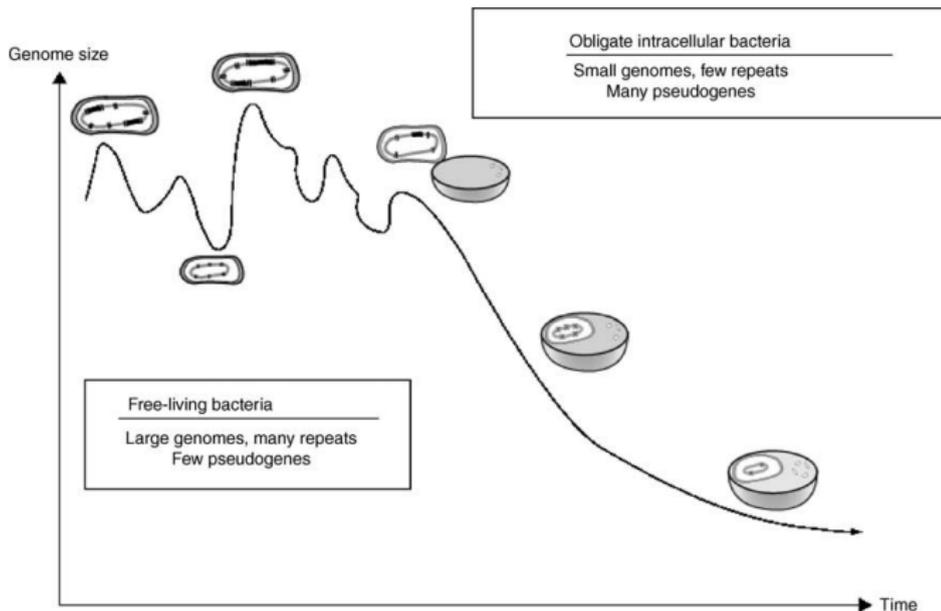


Figure 3. Schematic illustration of genome size variations as a function of time during transitions to intracellular growth habitats. Filled boxes represent mobile genetic elements. Genomes of obligate intracellular bacteria are smaller and have a lower content of repeated sequences (//) and a higher content of pseudogenes (x) than genomes of free-living bacteria and facultative intracellular parasites.

Genome size polymorphism in *E. coli*

Distribution of Chromosome Length Variation in Natural Isolates of *Escherichia coli*

Ulfar Bergthorsson and Howard Ochman

Department of Biology, University of Rochester

Large-scale variation in chromosome size was analyzed in 35 natural isolates of *Escherichia coli* by physical mapping with a restriction enzyme whose sites are restricted to rDNA operons. Although the genetic maps and chromosome lengths of the laboratory strains *E. coli* K12 and *Salmonella enterica* sv. Typhimurium LT2 are highly congruent, chromosome lengths among natural strains of *E. coli* can differ by as much as 1 Mb, ranging from 4.5 to 5.5 Mb in length. This variation has been generated by multiple changes dispersed throughout the genome, and these alterations are correlated; i.e., additions to one portion of the chromosome are often accompanied by additions to other chromosomal regions. This pattern of variation is most probably the result of selection acting to maintain equal distances between the replication origin and terminus on each side of the circular chromosome. There is a large phylogenetic component to the observed size variation: natural isolates from certain subgroups of *E. coli* have consistently larger chromosomes, suggesting that much of the additional DNA in larger chromosomes is shared through common ancestry. There is no significant correlation between genome sizes and growth rates, which counters the view that the streamlining of bacterial genomes is a response to selection for faster growth rates in natural populations.

Bergthorsson, U. and Ochman H. (1998) *Mol. Biol. Evol.*, **15** :6-16.

The ECOR collection

TABLE 1. Standard reference strains and electromorph mobility profiles

No.	Strain ^a Previous designation ^a	Source		References	Group ^b	Enzyme ^c										
		Host (sex)	Location			MDH	6PG	ADK	PE2	GOT	IDH	PGI	ACO	MPI	G6P	ADH
1	RM74A	Human (F)	Iowa	8, 9, 10, 12, 13, 15, 16	I	2	6	4	5	3	2	4	7	3	2	1
2	STM1	Human (M)	New York	12, 15	I	2	6	4	5	3	2	4	7	3	2	1
3	WIR1(a)	Dog	Massachusetts	12, 15	I	2	6	4	5	3	2	4	7	3	2	1
4	RM39A	Human (F)	Iowa	8-10	I	2	15	4	7	3	2	4	6	3	2	1
5	RM60A	Human (F)	Iowa	8, 9, 12, 13, 15, 16	I	2	4	4	5	3	2	4	7	3	2	1
6	RM66C	Human (M)	Iowa	5, 6, 8, 9, 11-13, 15, 16	I	2	13	4	5	3	2	4	6	3	2	1
7	RM73C	Orangutan	Washington (zoo)	5, 8, 9, 12, 13, 15	I	2	5	4	7	3	2	4	7	3	1	1
8	RM77C (b)	Human (F)	Iowa	4, 7-9, 12, 13, 15, 16	I	2	9	4	5	3	2	4	7	3	2	1
9	FN98	Human (F)	Sweden	2, 12, 15, 16	I	2	9	4	5	3	2	4	7	3	2	1
10	ANI	Human (F)	New York	12, 15	I	2	9	4	5	3	2	4	7	3	2	1
11	C97	Human (F)	Sweden	2, 12, 15, 16	I	2	9	4	5	3	2	4	7	3	2	1
12	FN59	Human (F)	Sweden	2, 12, 15, 16	I	2	6	4	5	3	5	4	7	3	2	4
13	FN10	Human (F)	Sweden	2, 12, 15, 16	I	2	6	4	7	3	2	4	7	3	1	1

Ochman, H. and Selander, R.K. (1984) *J. Bacteriol.*, **157** :690-693.

Digestion of the *E. coli* chromosome with I-CeuI

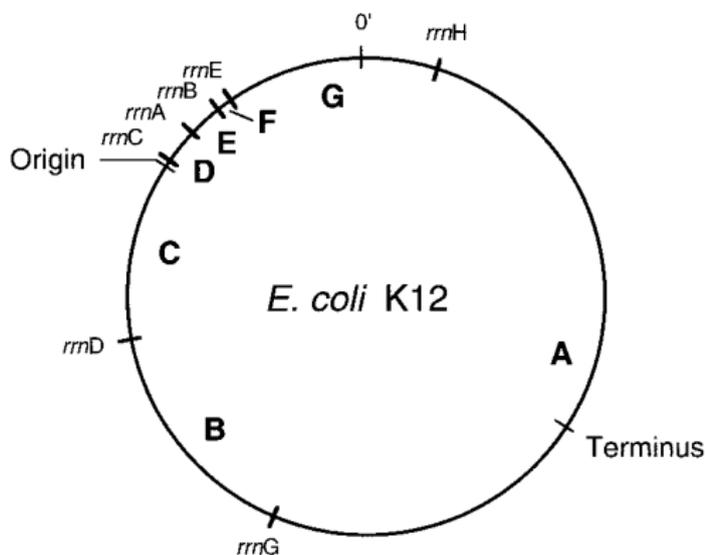


FIG. 1.—Locations of I-CeuI recognition sites on the *E. coli* K12 chromosome. I-CeuI cleaves at the seven *rrn* genes, whose map positions are indicated. The resulting restriction fragments are designated A through G.

Results in kb

	group	strain	Host..sex.	Location	A	B	C	D	E	F	G
1	A	ECOR4	Human (F)	Iowa	2585	707	527	90	166	38	608
2	A	ECOR5	Human (F)	Iowa	2940	743	515	90	128	38	699
3	A	ECOR11	Human (F)	Sweden	2750	824	556	90	128	38	735
4	A	ECOR13	Human (F)	Sweden	2485	680	515	90	128	38	639
5	A	ECOR14	Human (F)	Sweden	2645	735	608	90	128	38	707
6	A	ECOR15	Human (F)	Sweden	2690	735	575	90	138	38	639
7	A	ECOR18	Celebese ape	Washington	2510	699	515	90	122	38	608
8	A	ECOR19	Celebese ape	Washington	2480	699	527	90	122	38	639
9	A	ECOR20	Steer	Bali	2505	654	480	90	122	38	608
10	A	ECOR21	Steer	Bali	2505	654	480	90	122	38	608
11	A	ECOR23	Elephant	Washington	2675	807	532	90	138	38	680
12	B1	ECOR27	Giraffe	Washington	2600	707	515	90	143	38	616
13	B1	ECOR28	Human (F)	Iowa	2620	743	527	94	128	38	639
14	B1	ECOR29	Kangaroo rat	Nevada	2610	787	527	94	138	38	639
15	B1	ECOR34	Dog	Massachusetts	2500	790	515	94	138	38	680
16	B1	ECOR58	Lion	Washington	2700	743	515	94	136	38	639
17	B1	ECOR68	Giraffe	Washington	2745	843	532	94	138	38	807
18	B1	ECOR71	Human (F)	Sweden	2650	771	547	90	138	38	654
19	B1	ECOR72	Human (F)	Sweden	2635	771	532	94	138	38	680
20	B2	ECOR51	Human infant	Massachusetts	2750	810	550	112	138	38	810
...											
31	D	ECOR39	Human (F)	Sweden	2780	787	581	104	143	38	713
32	D	ECOR40	Human (F)	Sweden	2845	807	616	104	143	43	787
33	E	ECOR31	Leopard	Washington	2775	743	547	94	138	38	735
34	E	ECOR37	Marmoset	Washington	3100	787	581	94	175	38	743
35	E	ECOR42	Human (M)	Massachusetts	2735	743	616	94	143	38	699

What is the polymorphism of *E. coli* genome size ?

Study this yourself :

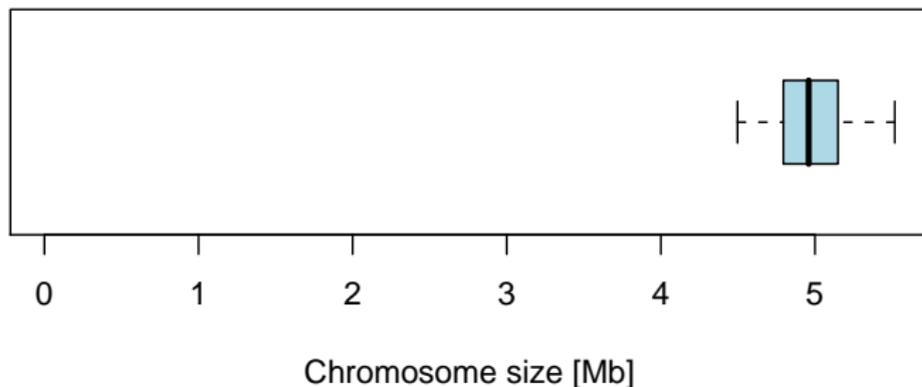
```
pgs <- read.table("http://pbil.univ-lyon1.fr/R/donnees/polygensize.txt",
  header = TRUE, sep = "\t")
head(pgs)
```

	subgroup	strain	Host..sex.	Location	A	B	C	D	E	F	G
1	A	ECOR4	Human (F)	Iowa	2585	707	527	90	166	38	608
2	A	ECOR5	Human (F)	Iowa	2940	743	515	90	128	38	699
3	A	ECOR11	Human (F)	Sweden	2750	824	556	90	128	38	735
4	A	ECOR13	Human (F)	Sweden	2485	680	515	90	128	38	639
5	A	ECOR14	Human (F)	Sweden	2645	735	608	90	128	38	707
6	A	ECOR15	Human (F)	Sweden	2690	735	575	90	138	38	639

- What is the distribution of genome size ?
- Any relationship with the subgroup ?
- What is the nice hidden structure in this dataset ?

Genome size is highly polymorphic in *E. coli*

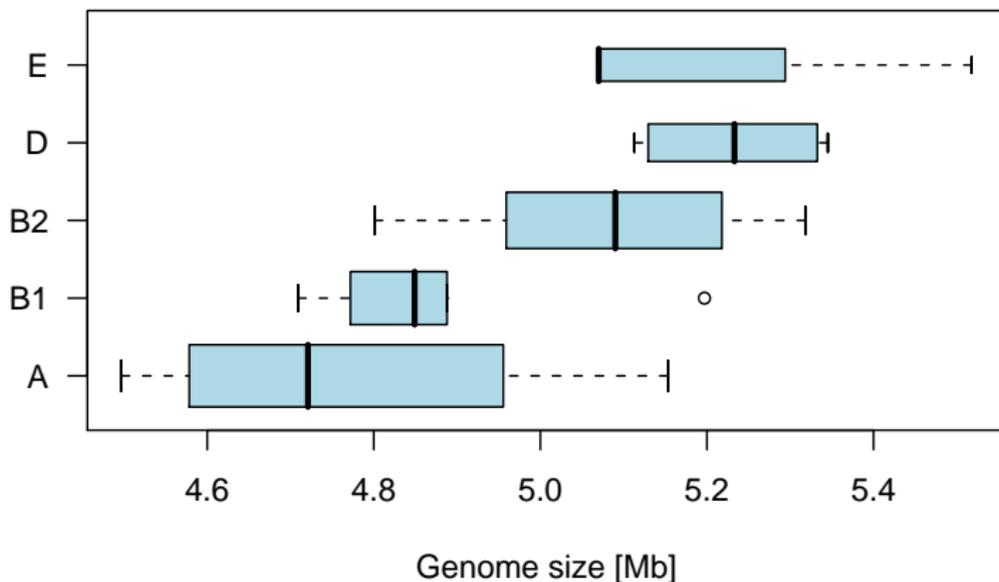
Distribution of genome size for 35 *Escherichia coli* strains



There is no meiotic constraints on chromosome length in bacteria.

Genome size phylogenetic inertia

Genome size within 5 subgroups of *Escherichia coli* strains



Genome size phylogenetic inertia

```
tcs <- rowSums(pgs[, 5:11])/1000
anova(lm(tcs ~ pgs$subgroup))
```

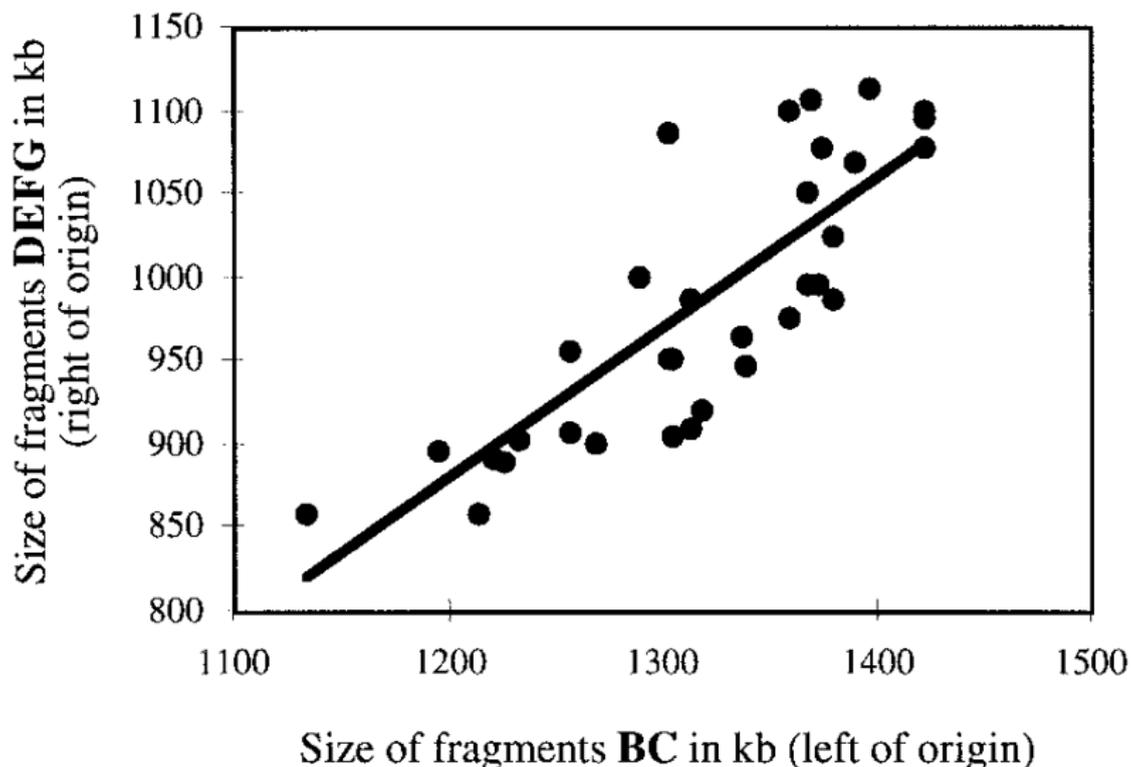
Analysis of Variance Table

Response: tcs

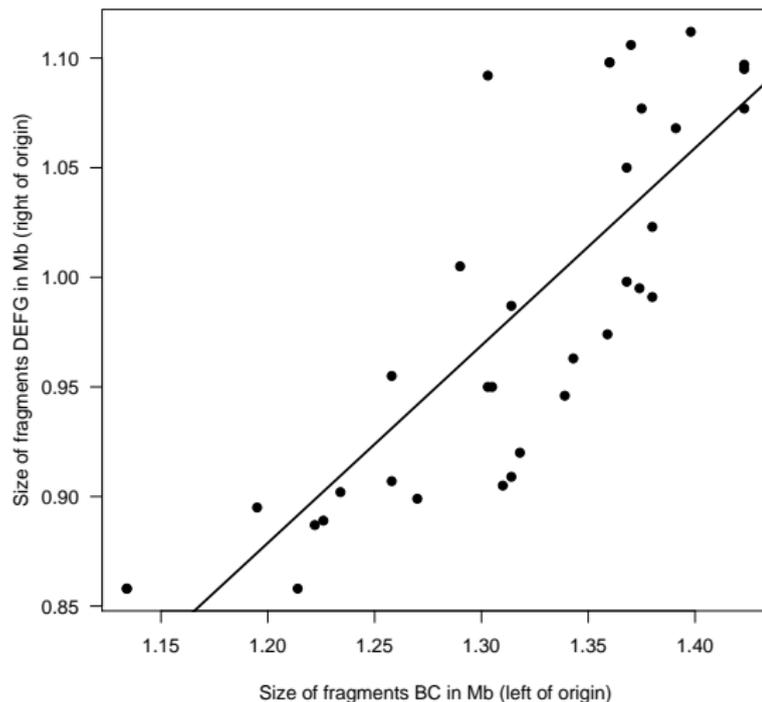
	Df	Sum Sq	Mean Sq	F value	Pr(>F)
pgs\$subgroup	4	1.07556	0.26889	6.817	0.0004999 ***
Residuals	30	1.18333	0.03944		

 Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

The nice hidden structure



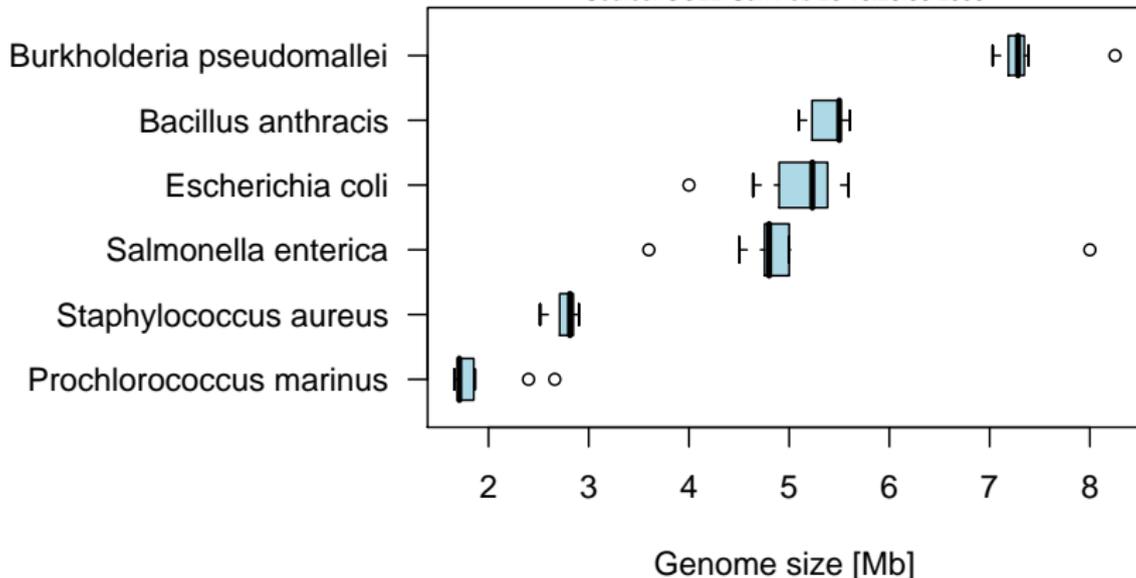
The nice hidden structure



Genome size polymorphism in bacteria

Genome size within species with at least 10 strains

Source: GOLD Sun Feb 26 16:28:56 2006

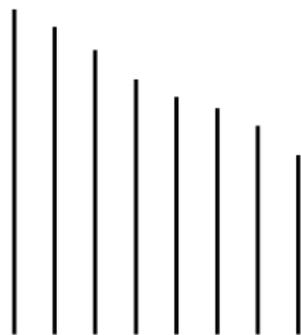


Topology

- 1 Introduction
- 2 Genome size
- 3 Topology & #**
- 4 G+C content
- 5 Replichores
- 6 Gene orientation biases
- 7 Chirochores
- 8 X-rated structure

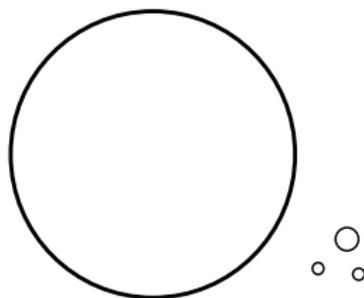
The big picture

Eukaryota



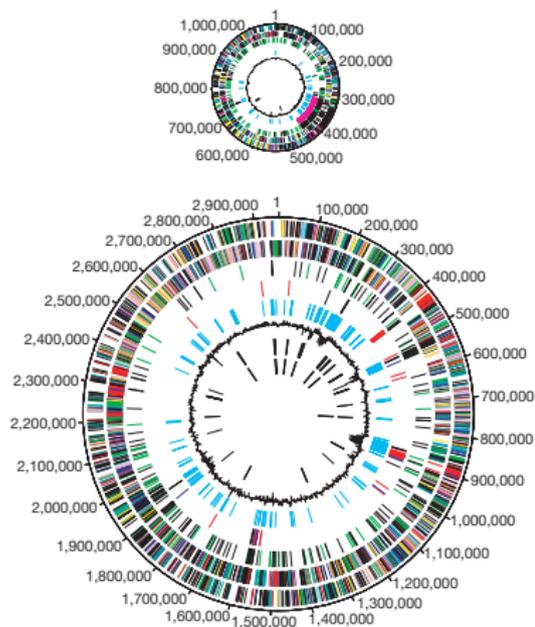
Many linear
chromosomes

Bacteria

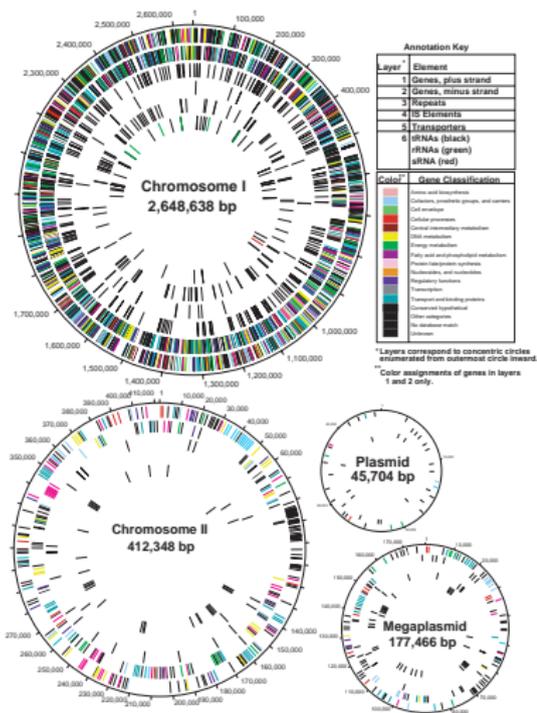


A single circular
chromosome + plasmids

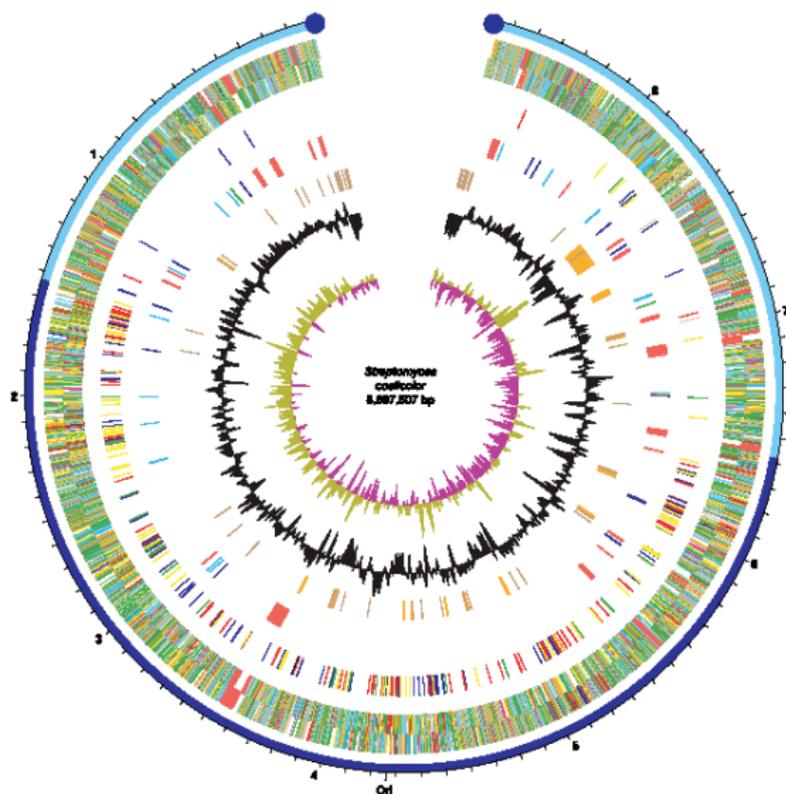
V. cholerae : 2 circular chromosomes



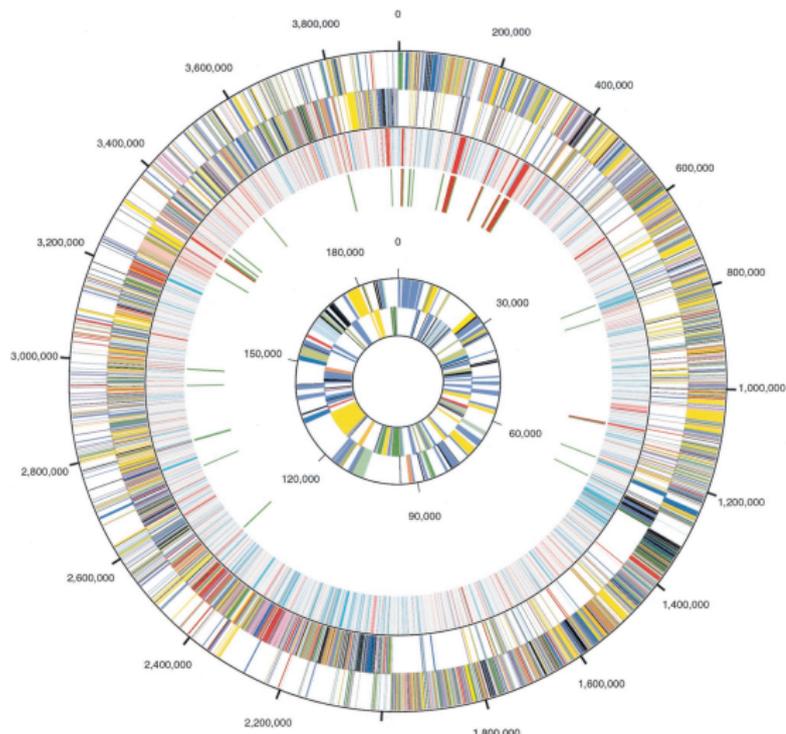
D. radiodurans : 2 circular chromosomes + 2 circular plasmids



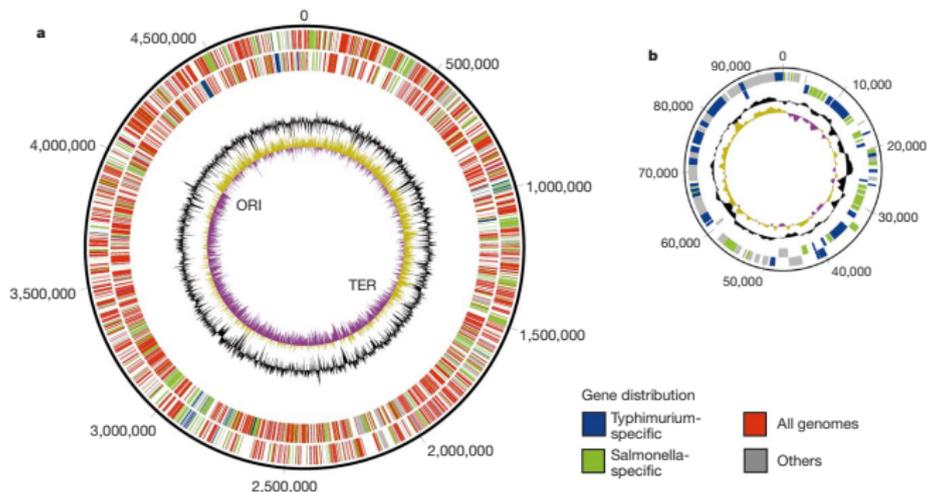
S. coelicolor : 1 linear chromosome



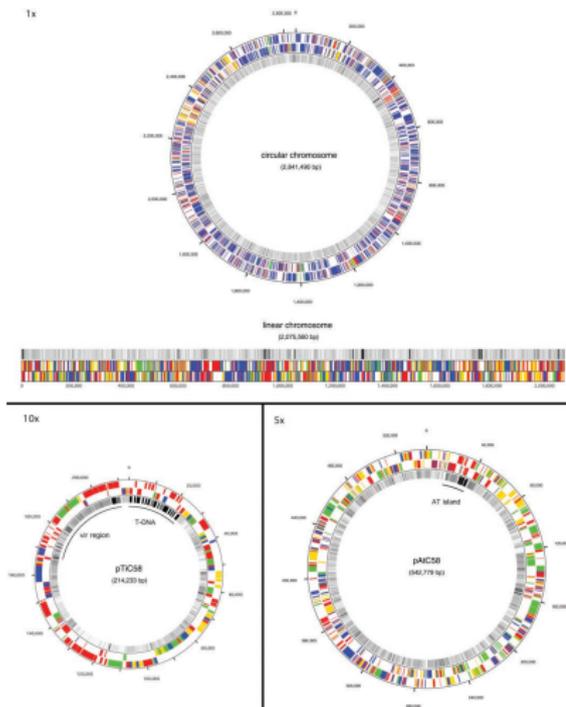
C. acetobutylicum : 1 circular chromosome + 1 circular megaplasmid



S. enterica serovar Typhimurium LT2 : 1 circular chromosome + 1 circular plasmid



A. tumefaciens : 1 circular chromosome + 1 linear chromosome + 2 circular plasmid

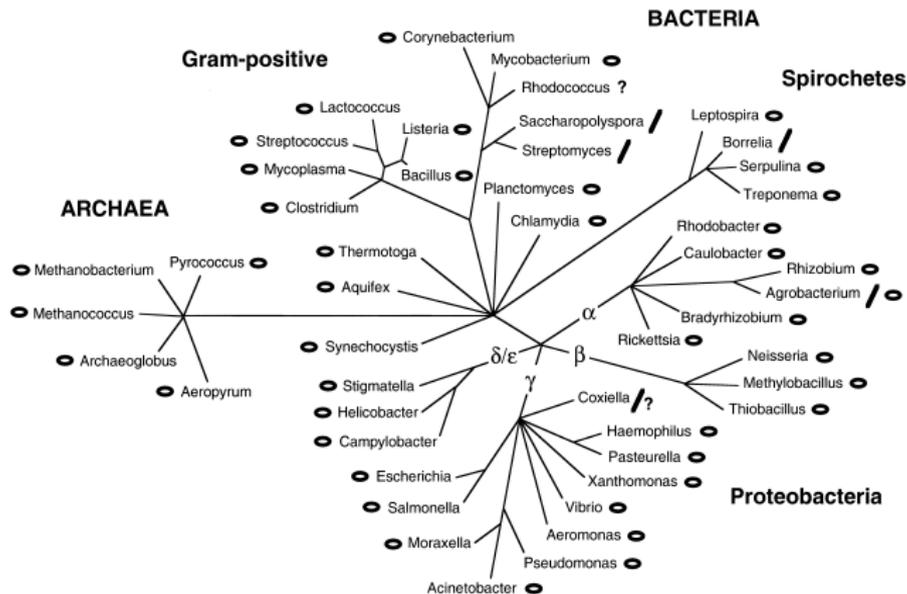


B. burgdorferi : many things !

Replicon	Geometry	Size (bp)
Chromosome ^h	Linear	910 725
cp9	Circular	9386
cp26	Circular	26 498
cp32-1	Circular	30 750
cp32-3	Circular	30 223
cp32-4	Circular	30 299
cp32-6	Circular	29 838
cp32-7	Circular	30 800
cp32-8	Circular	30 885
cp32-9	Circular	30 651
lp5 ⁱ	Linear	5228
lp17 ⁱ	Linear	16 928 ^k
lp21 ⁱ	Linear	18 901
lp25 ⁱ	Linear	24 177
lp28-1 ⁱ	Linear	28 250 ^k
lp28-2	Linear	29 766
lp28-3 ⁱ	Linear	28 601
lp28-4 ⁱ	Linear	27 323
lp36 ^j	Linear	36 849
lp38 ⁱ	Linear	38 829
lp54	Linear	53 541
lp56 (cp32) ^j	Linear	30 349
lp56 (other) ^{i,j}	Linear	22 622 ^k
Pseudogene plasmid ⁱ total		247 708
Other plasmid total		362 986
All plasmid total		610 694

Circular → linear

144

J.-N. Voff, J. Altenbuchner / FEMS Microbiology Letters 186 (2000) 143–150

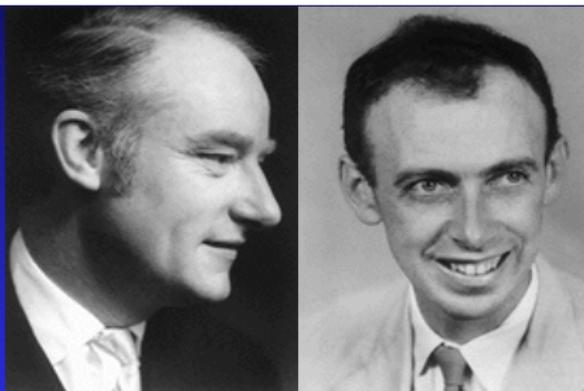
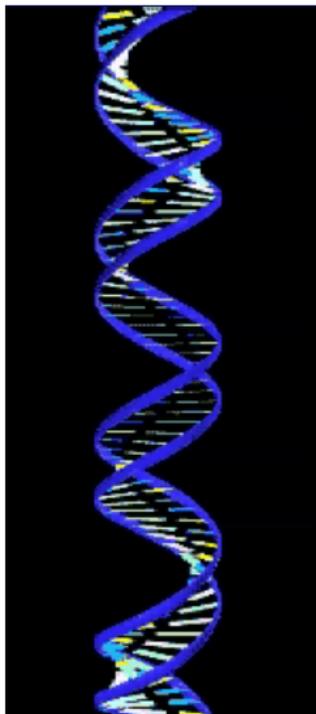
G+C content

- 1 Introduction
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G+C content

- Is calculated in percentage of G+C : $100 \frac{[G+C]}{[A+T+C+G]}$
- First nucleic acid technology applied to bacterial systematics
- One of the genomic characteristics recommended for the description of species and genera
- 5% and 10% are the common range found within a species and a genera, respectively
- Modulates the aminoacid content of proteins
- Source of troubles for phylogenetic inference

DNA double helix



Watson and Crick, Nature, 1953

The original figure



This figure is purely diagrammatic. The two ribbons symbolize the two phosphate-sugar chains, and the horizontal rods the pairs of bases holding the chains together. The vertical line marks the fibre axis

G+C content is the same for both strands

Let

$$A_a C_c G_g T_t$$

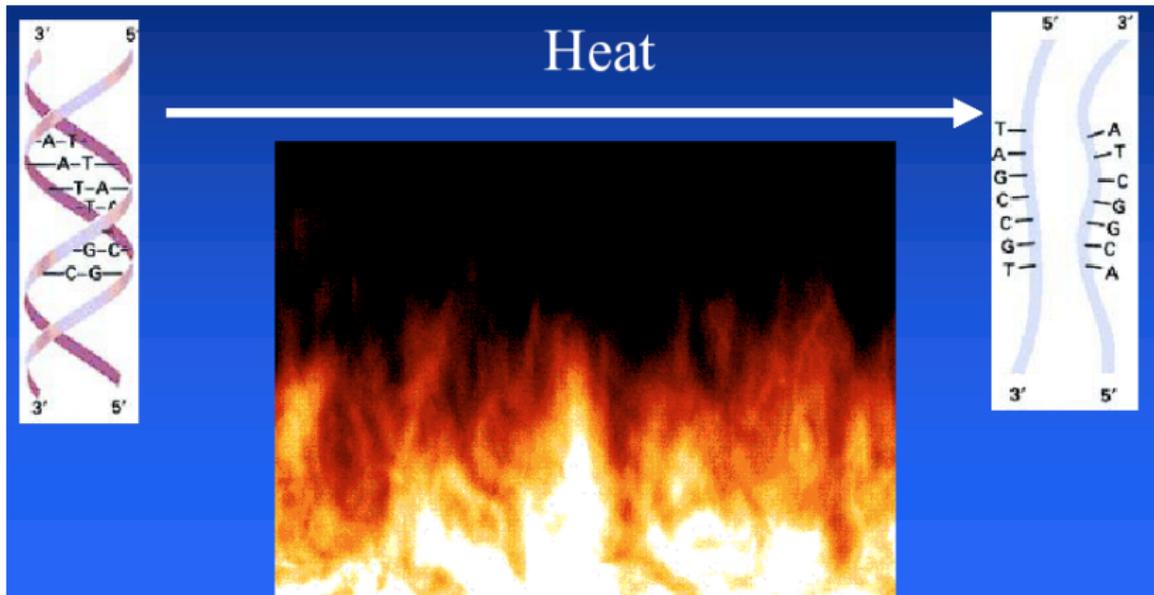
the primary formula on one strand. Then, its complementary strand composition is given by :

$$A_t C_g G_c T_a$$

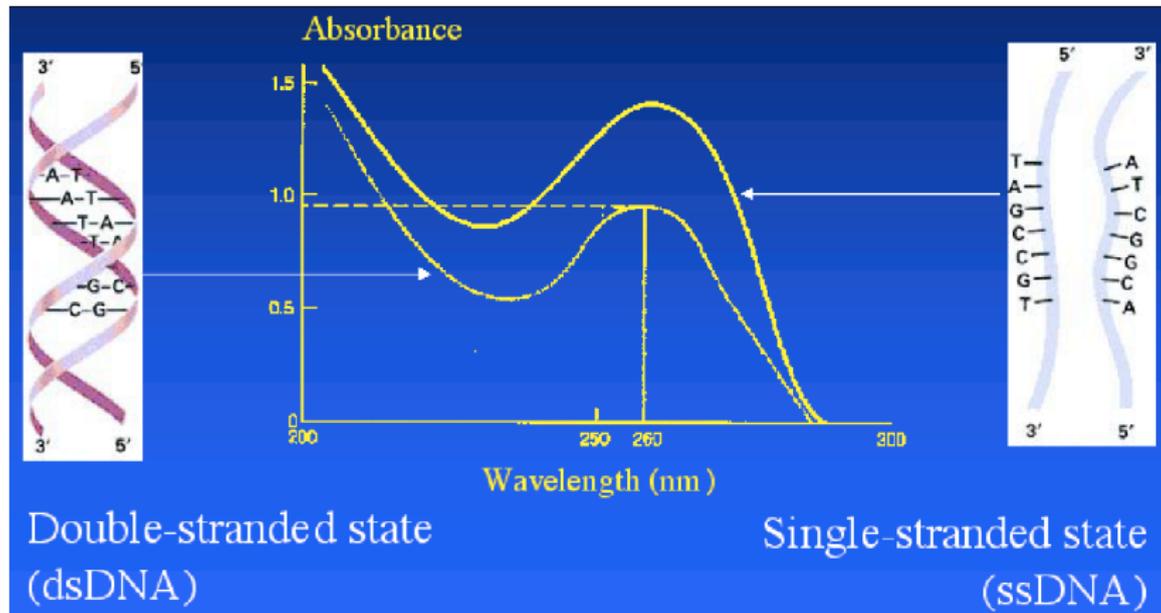
The G+C content is not affected :

$$\frac{g + c}{a + c + g + t} = \frac{c + g}{t + g + c + a}$$

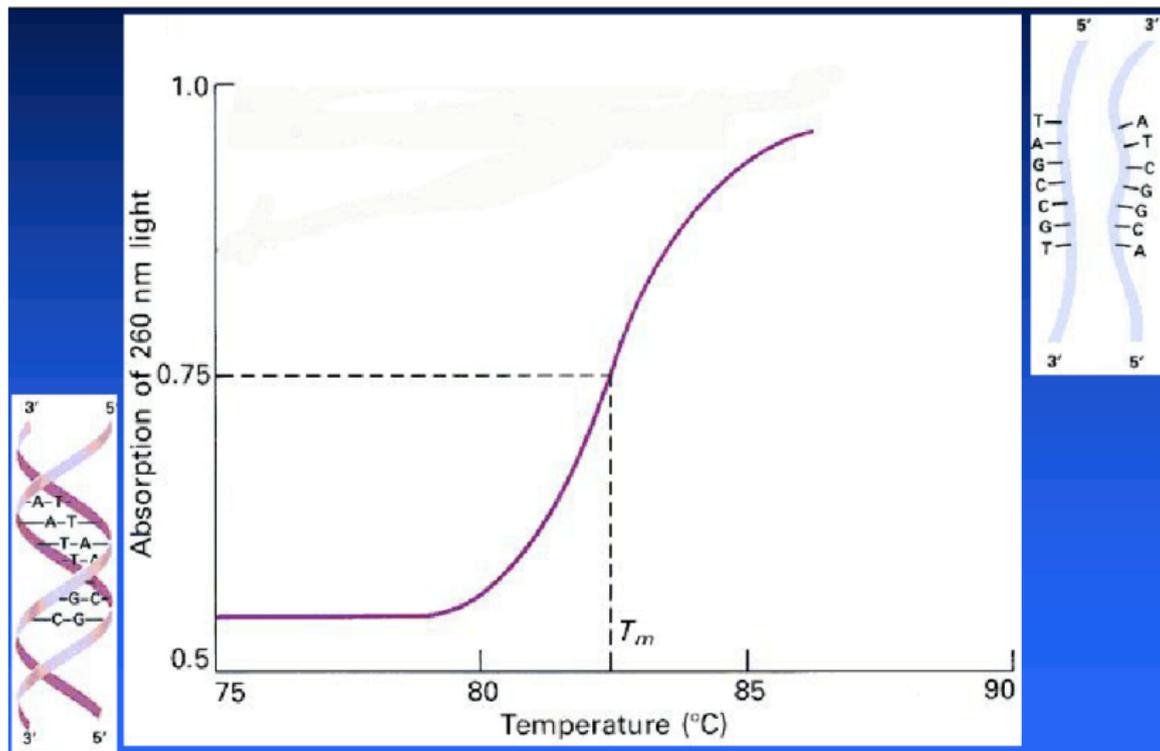
DNA denaturation



ssDNA & dsDNA absorbance

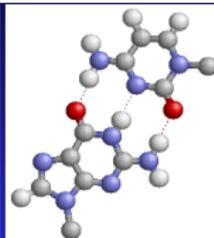
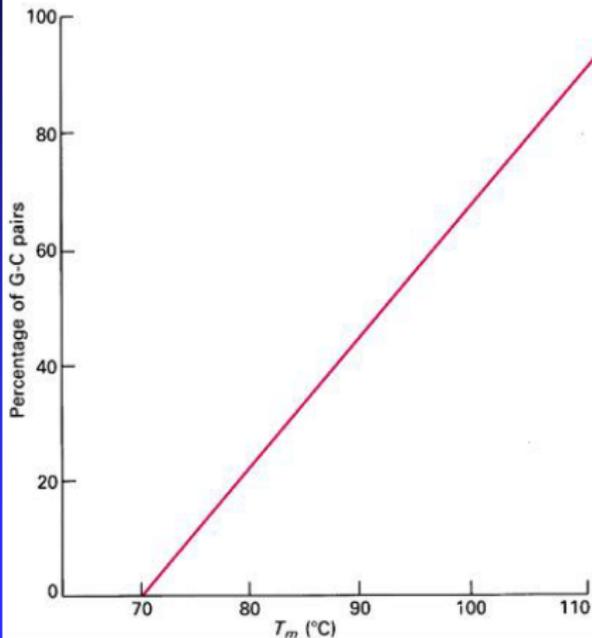
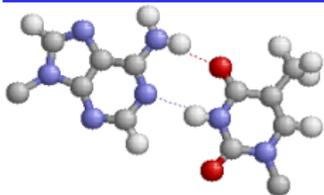


T_m is the temperature at midpoint of transition



T_m increases with DNA G+C content

AT pairs:
2 hydrogen
bonds



GC pairs:
3 hydrogen
bonds

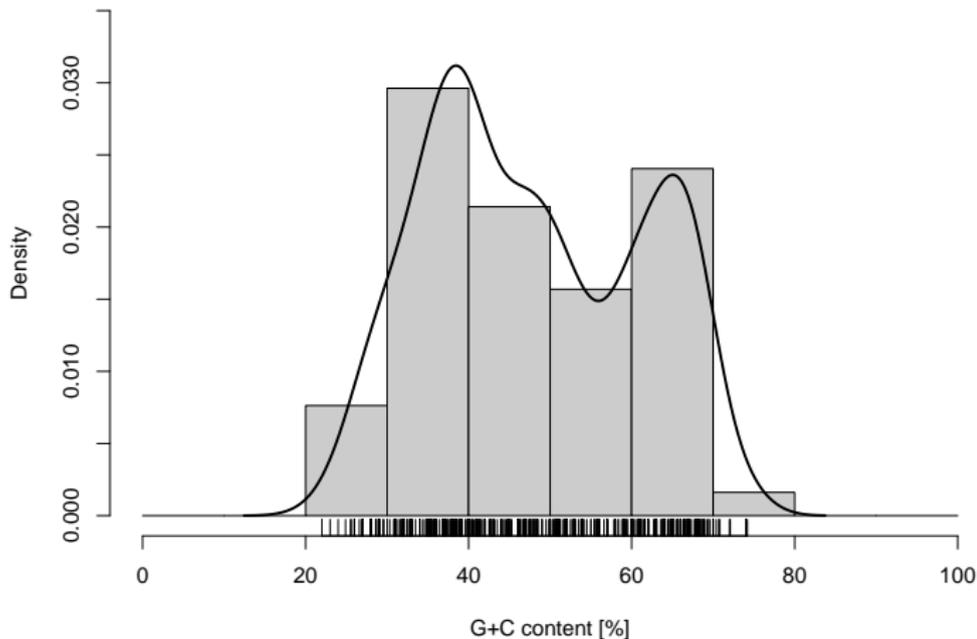
What is the distribution of G+C content in "bacteria"?

Study this yourself :

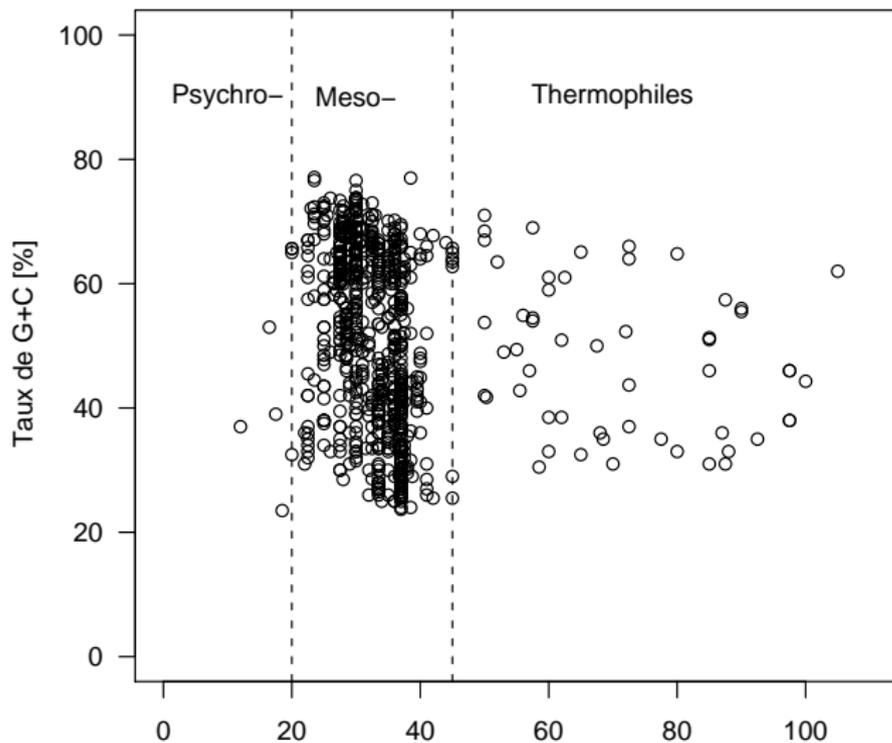
- 1 extract G+C data from GOLD
<http://www.genomesonline.org/> and study its distribution.
- 2 study data from ftp:
<ftp://pbil.univ-lyon1.fr/pub/datasets/JME97/species>.
What is the relationship with optimum growth temperature T_{opt} ?
- 3 study data from
[http://pbil.univ-lyon1.fr/R/donnees/gc02.txt](ftp://pbil.univ-lyon1.fr/R/donnees/gc02.txt). What is the relationship with (an)aerobiosis ?

G+C content from GOLD

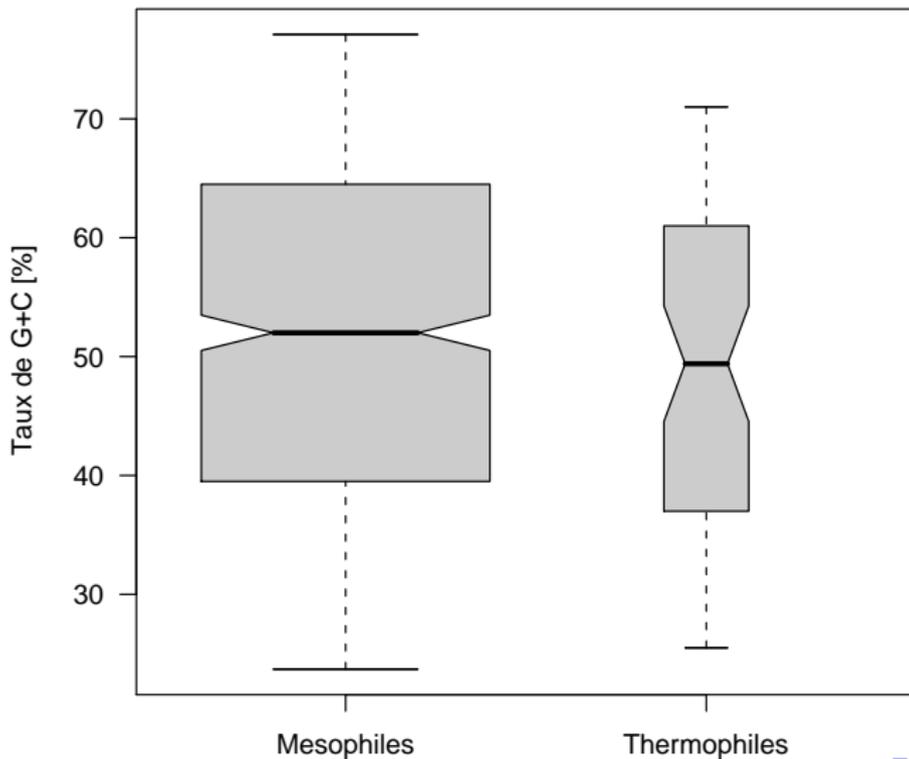
GOLD data 05-MAR-2006 (n = 682 strains)



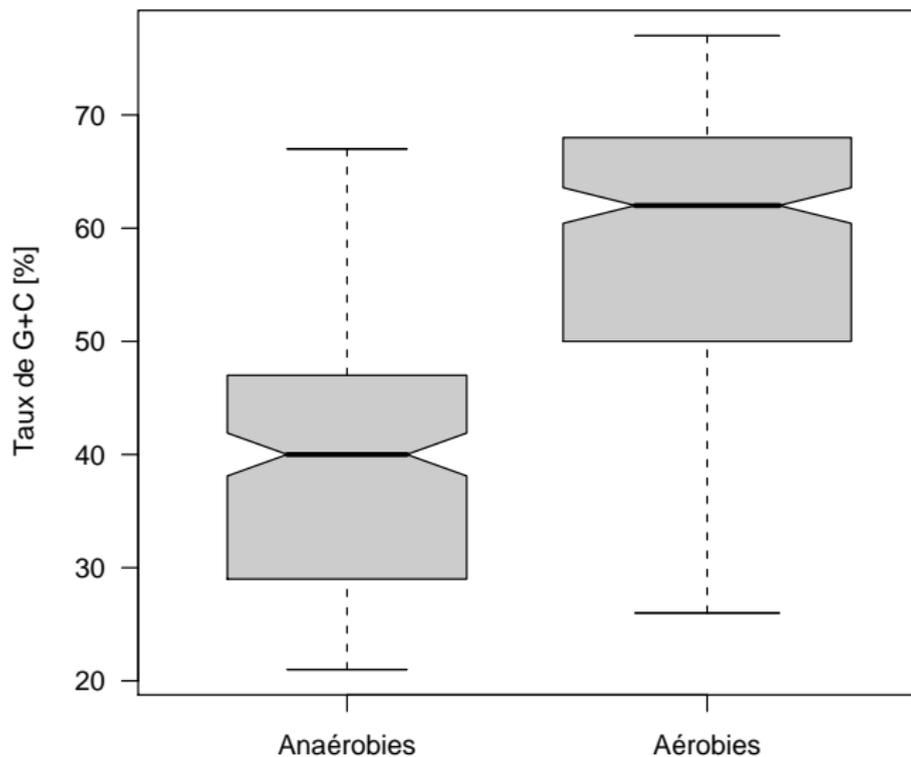
G+C content and T_{opt}



G+C content and T_{opt}



G+C content and aerobiosis



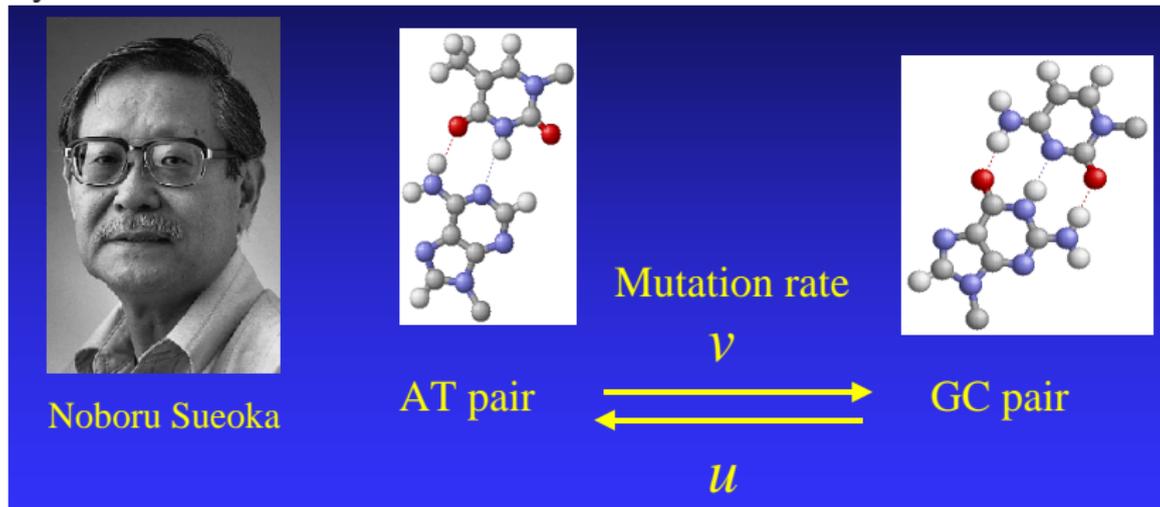
The distribution of G+C content in "bacteria"

Results from your study :

- 1 G+C content ranges from $\approx 25\%$ to 75% in "bacteria".
- 2 G+C content is not correlated with T_{opt} .
- 3 G+C content is correlated with aerobiosis : aerobic "bacteria" have a significant higher G+C than anerobic "bacteria".

Underlying mechanism

Symmetric Directional Mutation Pressure :



Sueoka, N. (1962) On the genetic basis of variation and heterogeneity of DNA base composition. *Proc. Natl. Acad. Sci. USA*, **48** :582-592

Underlying mechanism

Direct experimental evidence :

Cox, E.C., Yanofsky, C. (1967) Altered base ratios in the DNA of an *Escherichia coli* mutator strain. *Proc. Natl. Acad. Sci. USA*, **58** :1895-1902

Accelerated evolution experiment with a mutator strain : G+C content variation visible at a lab time scale.

Underlying mechanism

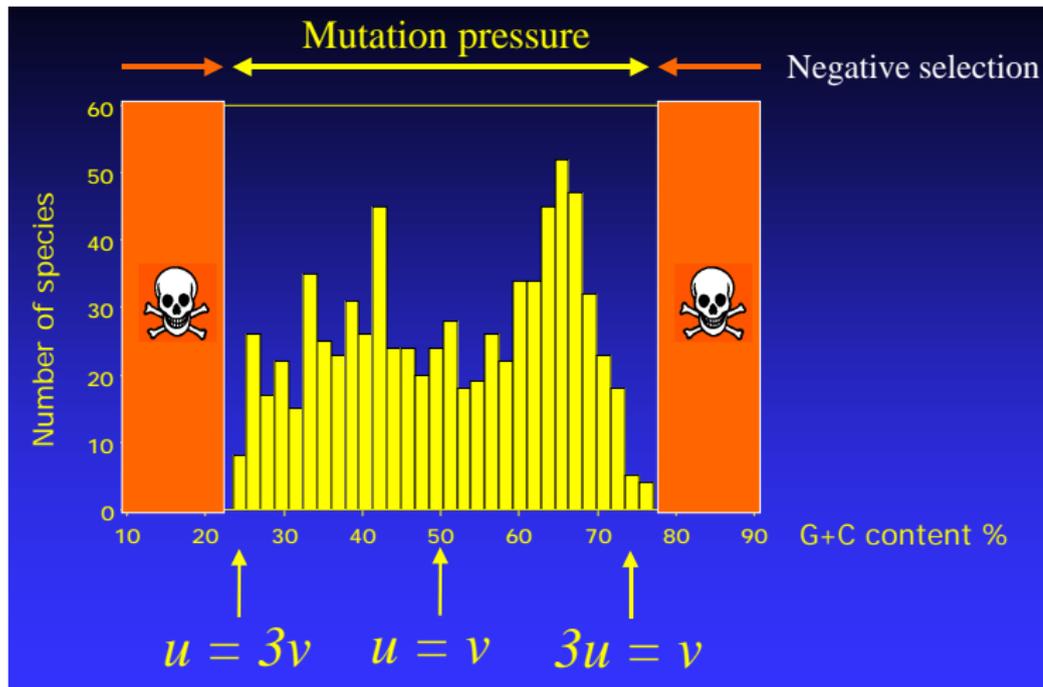
$$\frac{d\theta}{dt} = v(1 - \theta) - u\theta$$

$$\theta(t) = \left(\theta_0 - \frac{v}{u+v} \right) e^{-(u+v)t} + \frac{v}{u+v}$$

G+C content at equilibrium :

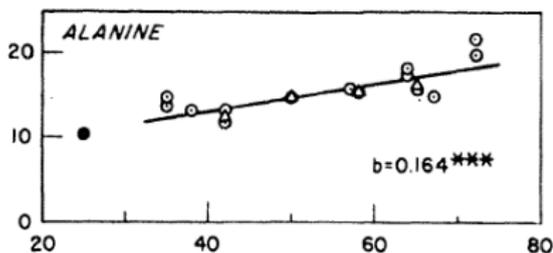
$$\theta^* = \theta(+\infty) = \frac{v}{u+v}$$

Underlying mechanism



G+C content and aa content

The impact of G+C content on the amino-acid composition of proteins was known even before the deciphering of the genetic code :



Sueoka, N. (1961) Correlation between base composition of deoxyribonucleic acid and amino acid composition of protein. *Proc. Natl. Acad. Sci. USA*, **48** :582-592

- Was used as a clue to crack the genetic code (e.g. here Ala codons are expected to be G+C rich, and yes indeed GCN codons are G+C rich!).
- First evidence that the genetic code is (almost) universal.

A null hypothesis for the aa content of proteins

- CDS are build by random sampling from an urn with a given G+C content θ .
- We assume for the shake of simplicity that $C = G = \frac{\theta}{2}$ and $A = T = \frac{1-\theta}{2}$.
- What would be the amino-acid composition of proteins under this simplistic model ?

A null hypothesis for the aa content of proteins

Let $X_i \in \{A, C, G, T\}$ a random variable for the result of outcome number i . Note $P_A = P(X_i = A)$, $P_C = P(X_i = C)$, $P_G = P(X_i = G)$, $P_T = P(X_i = T)$ the probabilities for the four bases. We have assumed that $P_C = P_G = \frac{\theta}{2}$ and $P_A = P_T = \frac{1-\theta}{2}$.

The probability for codon GAA is for instance :

$$P(GAA) = P(X_1 = G \cap X_2 = A \cap X_3 = A) = P_G P_A P_A$$

In coding sequences there are no stop codons (TAA, TAG or TGA)

so that :

$$P(GAA|\text{not - stop}) = \frac{P(GAA)}{P(\text{not-stop})} = \frac{P_G P_A P_A}{1 - (P_T P_A P_A + P_T P_A P_G + P_T P_G P_A)}$$

A null hypothesis for the aa content of proteins

At the amino-acid level, Glu is encoded by GAA or GAG, so that :

$$P(\text{Glu}) = P(\text{GAA} \cup \text{GAG} | \text{not - stop}) =$$

$$P(\text{GAA} | \text{not - stop}) + P(\text{GAG} | \text{not - stop})$$

In a similar way, we can deduce of the expected frequencies for all amino-acids under the model.

A null hypothesis for the aa content of proteins

$$P(\theta, aa) = \frac{f(\theta)}{8 - (1 - \theta)^2(1 + \theta)}$$

$$f(\theta) = \begin{cases} (1 - \theta)^2(2 - \theta) & \text{si } aa \in \{\text{Ile}\} \\ (1 - \theta)^2 & \text{si } aa \in \{\text{Phe, Lys, Tyr, Asn}\} \\ 1 - \theta^2 & \text{si } aa \in \{\text{Leu}\} \\ (1 - \theta)^2\theta & \text{si } aa \in \{\text{Met}\} \\ (1 - \theta)\theta & \text{si } aa \in \{\text{Asp, Glu, His, Gln, Cys}\} \\ 2(1 - \theta)\theta & \text{si } aa \in \{\text{Val, Thr}\} \\ 3(1 - \theta)\theta & \text{si } aa \in \{\text{Ser}\} \\ (1 - \theta)\theta^2 & \text{si } aa \in \{\text{Trp}\} \\ \theta(\theta + 1) & \text{si } aa \in \{\text{Arg}\} \\ 2\theta^2 & \text{si } aa \in \{\text{Gly, Pro, Ala}\} \end{cases}$$

What is the impact of G+C on the aa content ?

Study this yourself :

<ftp://pbil.univ-lyon1.fr/pub/datasets/GENE97/README>

- Differentiate cytoplasmic proteins and integral membrane proteins (by color or point style).
- Show the influence of G+C content for Ala, Lys, and Glu (at least).
- Add the null model as a line.

Replichores

- 1 Introduction
- 2 Genome size
- 3 Topology & #
- 4 G+C content
- 5 Replichores**
- 6 Gene orientation biases
- 7 Chirochores
- 8 X-rated structure

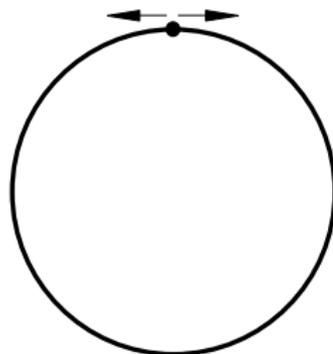
Replichores : origin and terminus of replication

Eukaryota



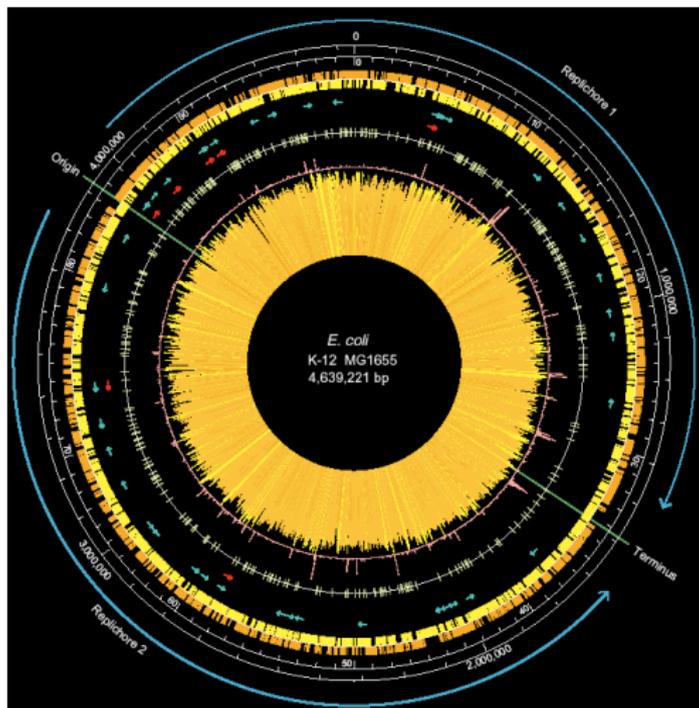
Many origins

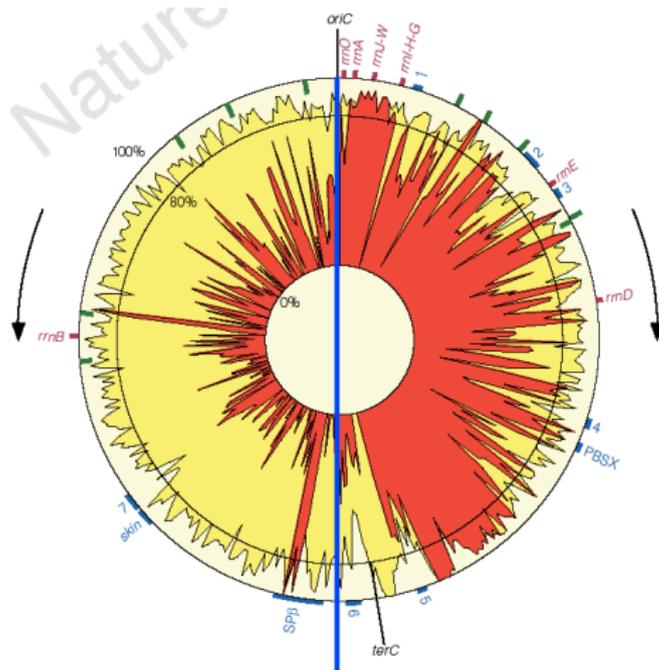
Bacteria



A single origin

There are two replichores per chromosome

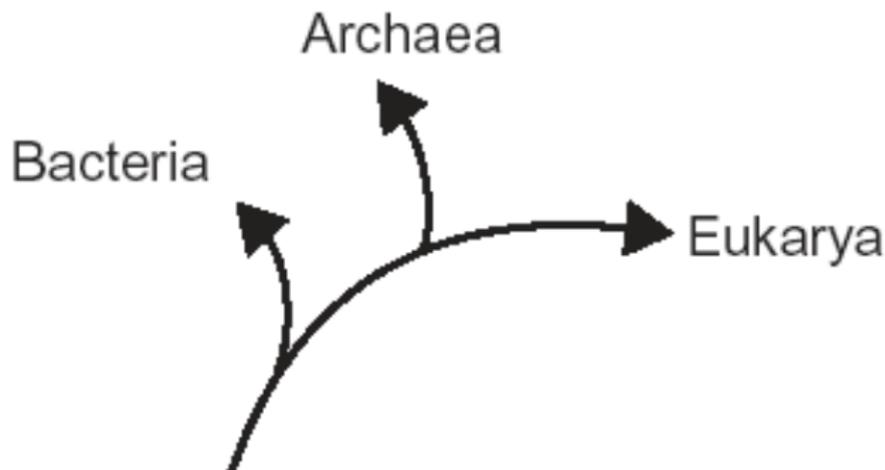


$\approx \pi$ 

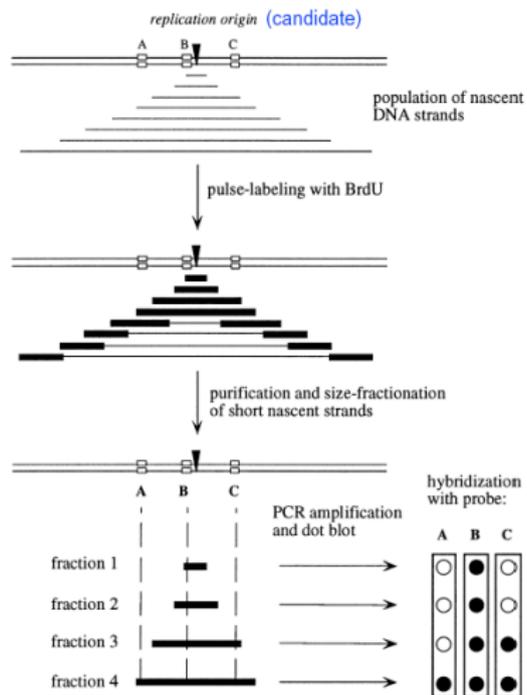
Archae & Bacteria

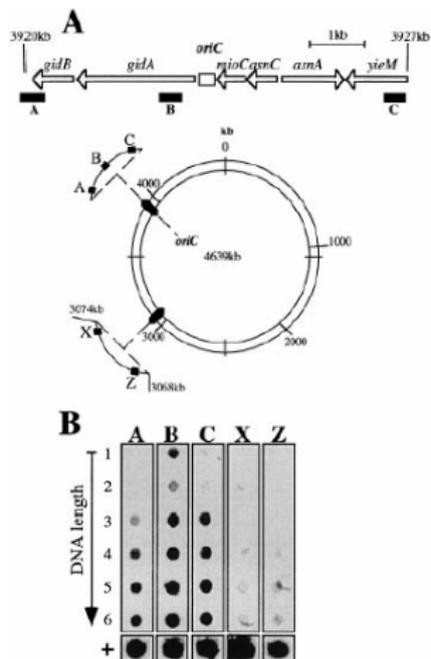
Small circular genome
with a single origin of
replication

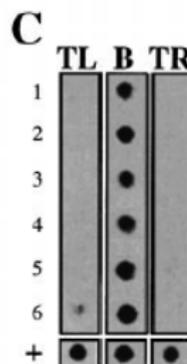
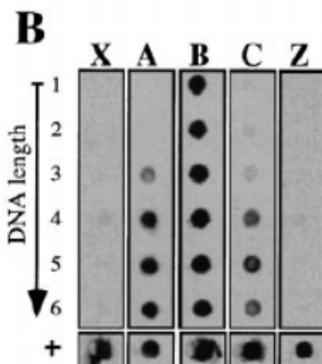
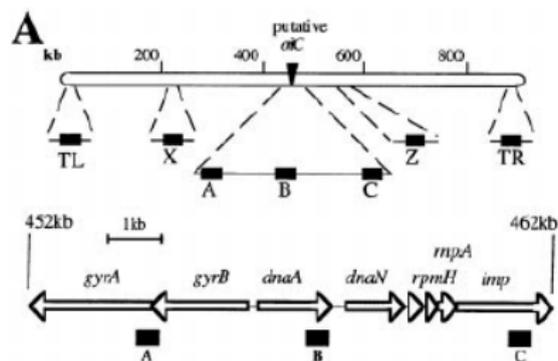
Similar replication factors



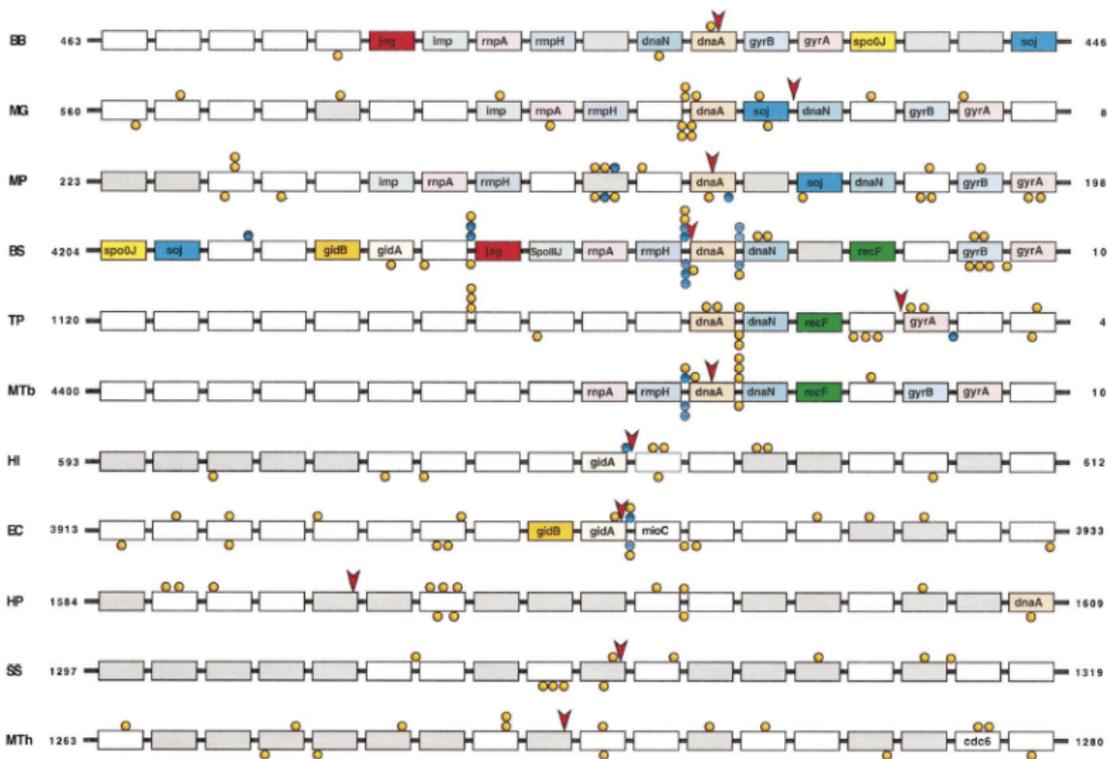
Looking for the origin



Looking for the origin in *E. coli*

Looking for the origin in *B. burgdorferi*

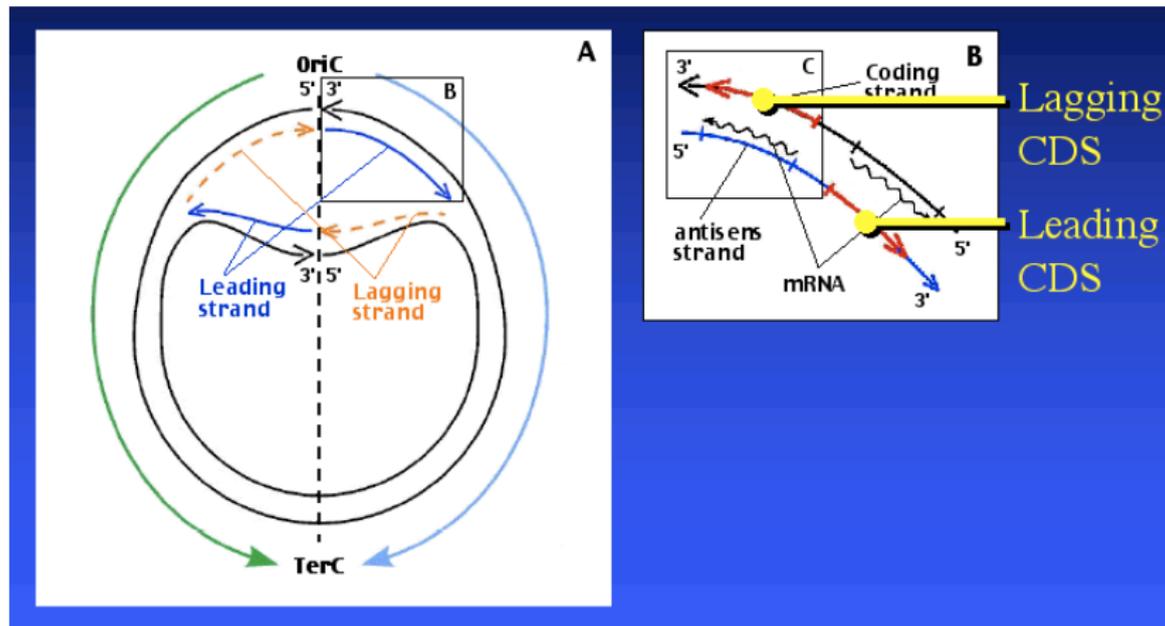
Zoom at the origin



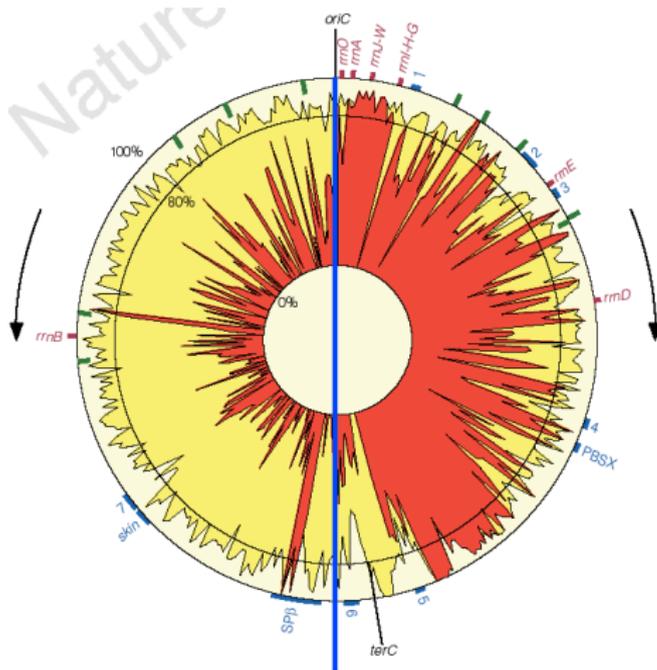
Gene orientation biases

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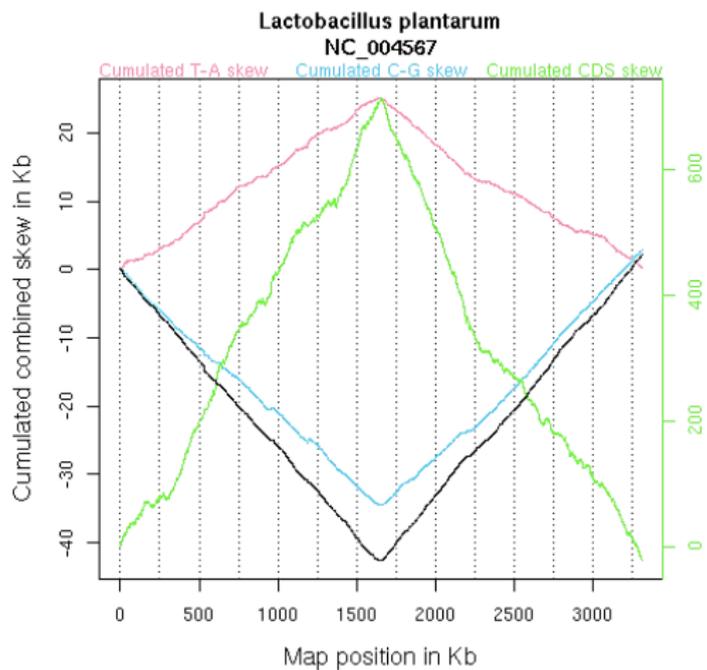
Leading and lagging CDS



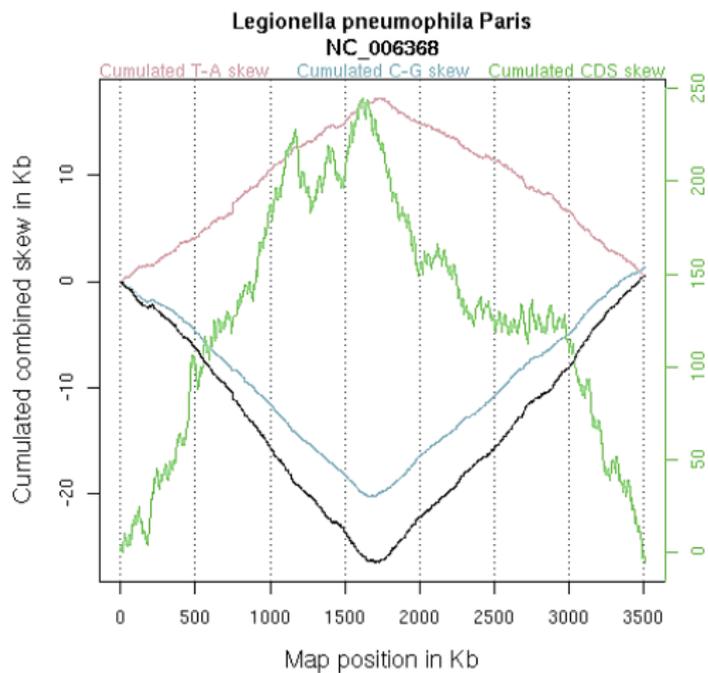
More leading CDS than lagging CDS



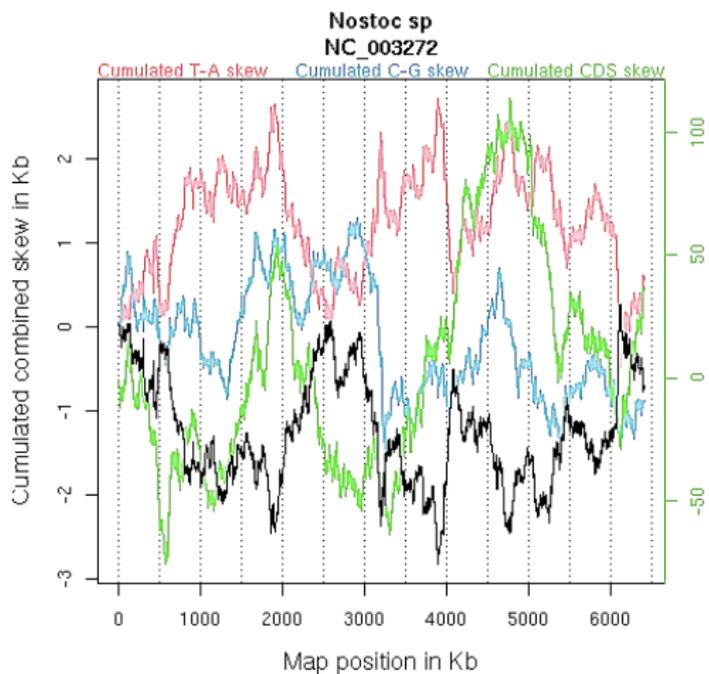
Lactobacillus plantarum



Legionella pneumophila



Nostoc sp



Collisions between polymerases

1: Science 1992 Nov 20;258(5086):1362-5

Related Articles, Books

Consequences of replication fork movement through transcription units in vivo.

French S.

Department of Biology, University of Virginia, Charlottesville 22903.

To examine the basis for the evolutionary selection for codirectionality of replication and transcription in *Escherichia coli*, electron microscopy was used to visualize replication from an inducible ColE1 replication origin inserted into the *Escherichia coli* chromosome upstream (5') or downstream (3') of *rnmB*, a ribosomal RNA operon. Active *rnmB* operons were replicated either in the same direction in which they were transcribed or in the opposite direction. In either direction, RNA polymerases were dislodged during replication. When replication and transcription were codirectional, the rate of replication fork movement was similar to that observed in nontranscribed regions. When replication and transcription occurred in opposite directions, replication fork movement was reduced.

Connection with essentiality

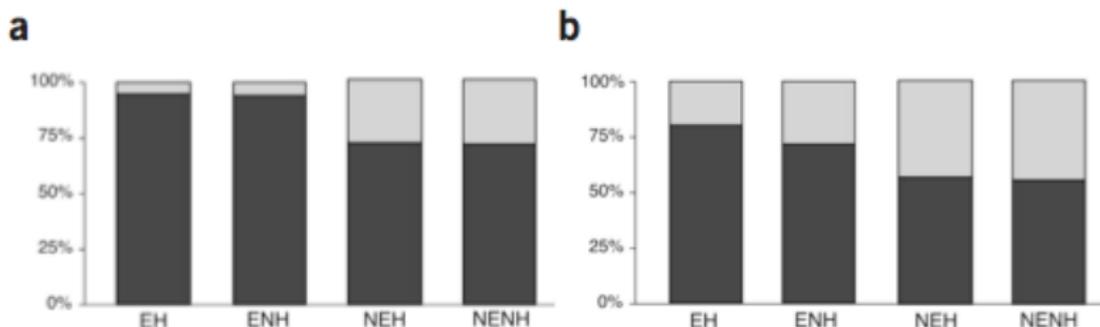
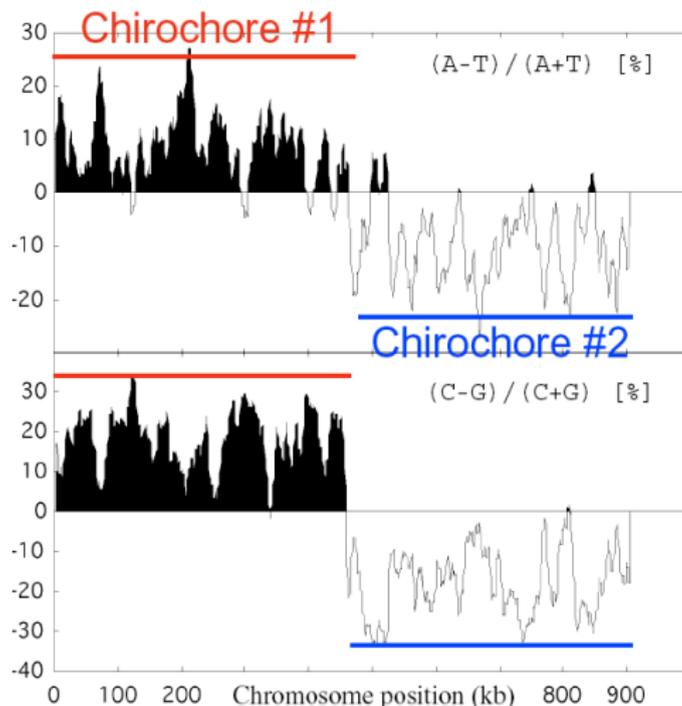


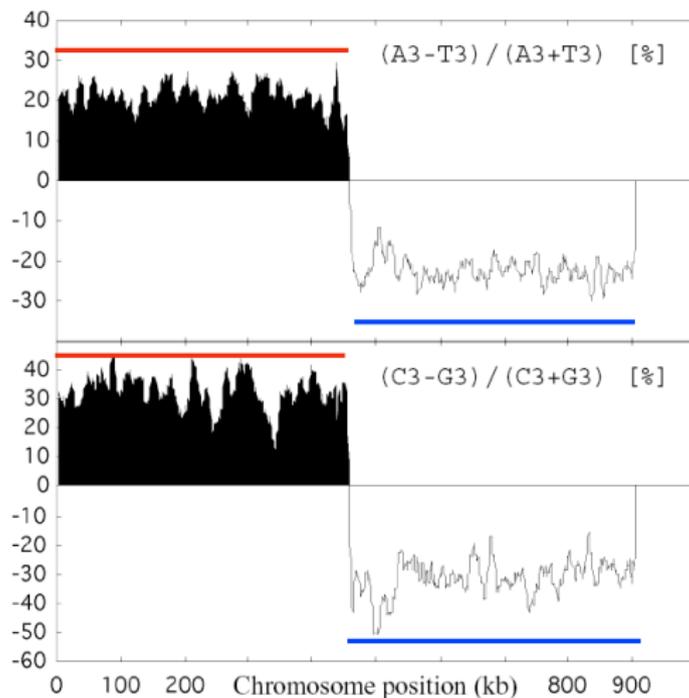
Figure 1 Distribution of genes between the leading (dark gray) and the lagging (light gray) strands of the genome of *B. subtilis* (a) and *E. coli* (b). H, highly expressed; NH, non-highly expressed; E, essential; NE, non-essential.

Rocha, E.P.C. & Danchin, A. (2003) *Nature Genetics*, **34** :377-378.

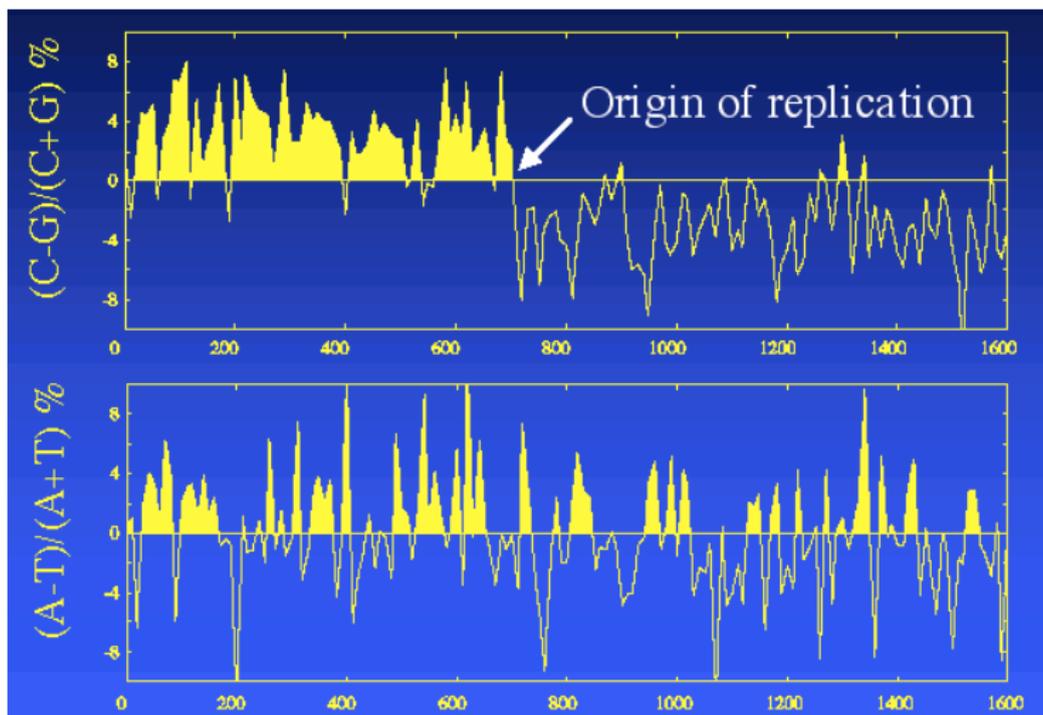
Chirochores : base composition biases

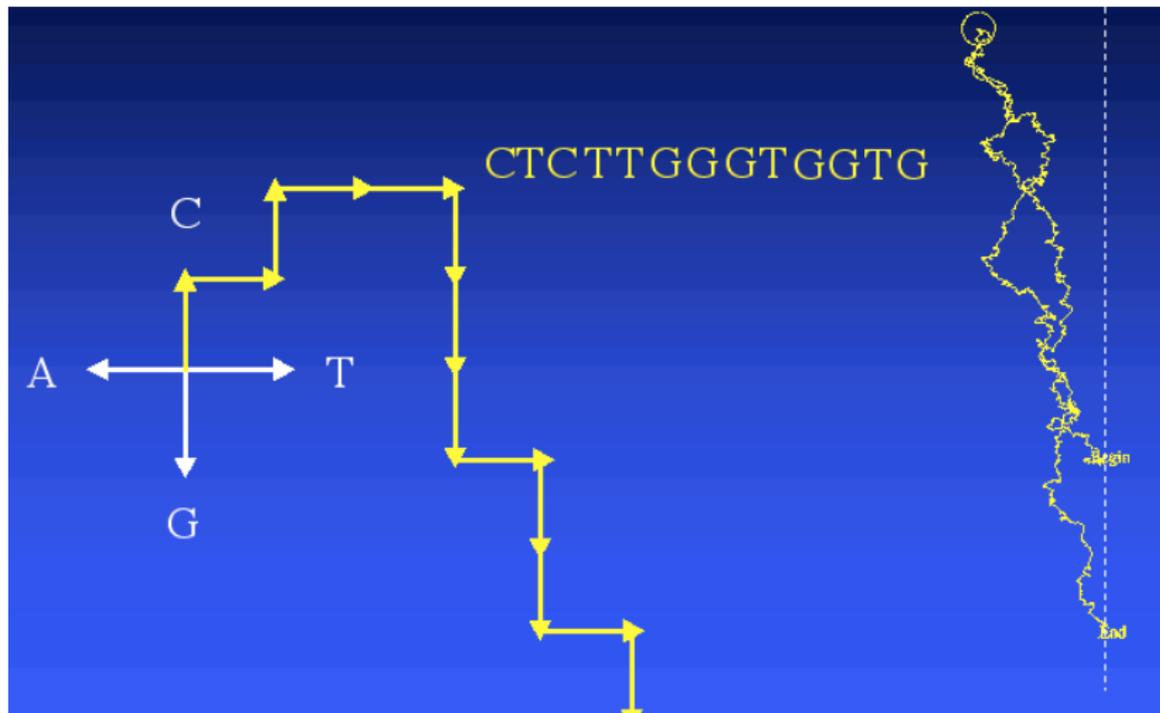
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- 5 Replichores
- 6 Gene orientation biases
- 7 Chirochores**
- 8 X-rated structure

Chirochores in *B. burgdorferi*

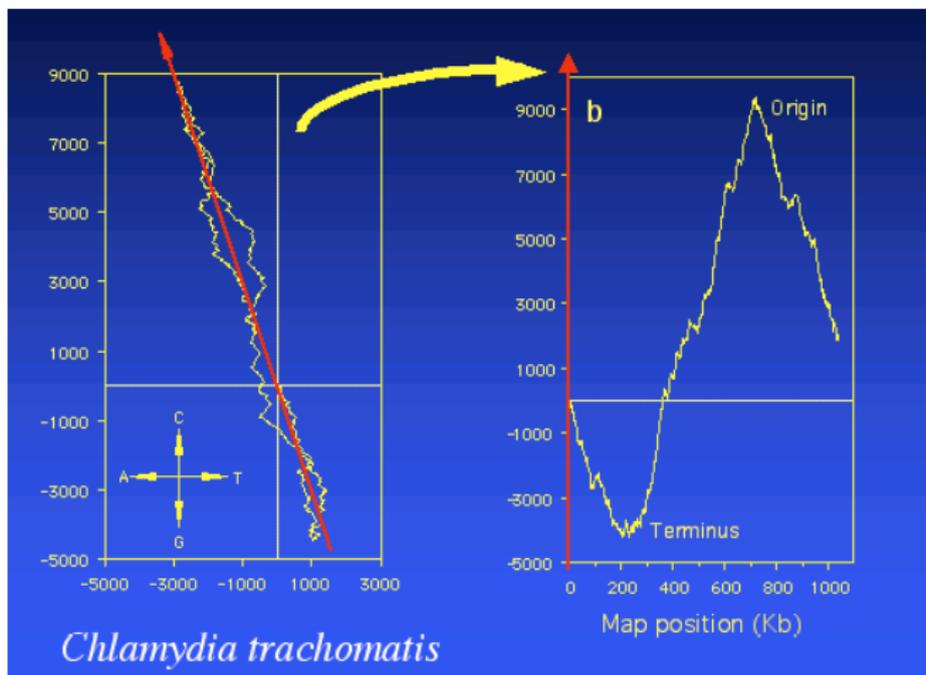
Chirochores in *B. burgdorferi* (third codon positions)

Usually GC skew $>$ AT skew e.g. *E. coli*

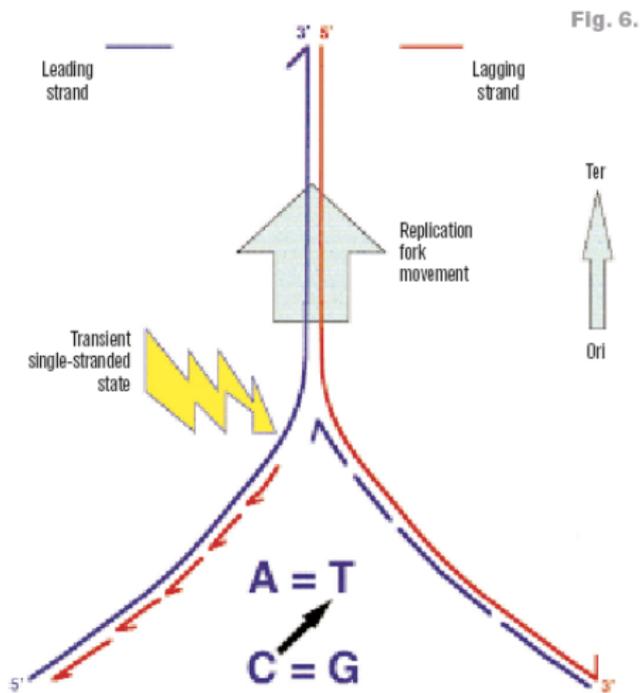


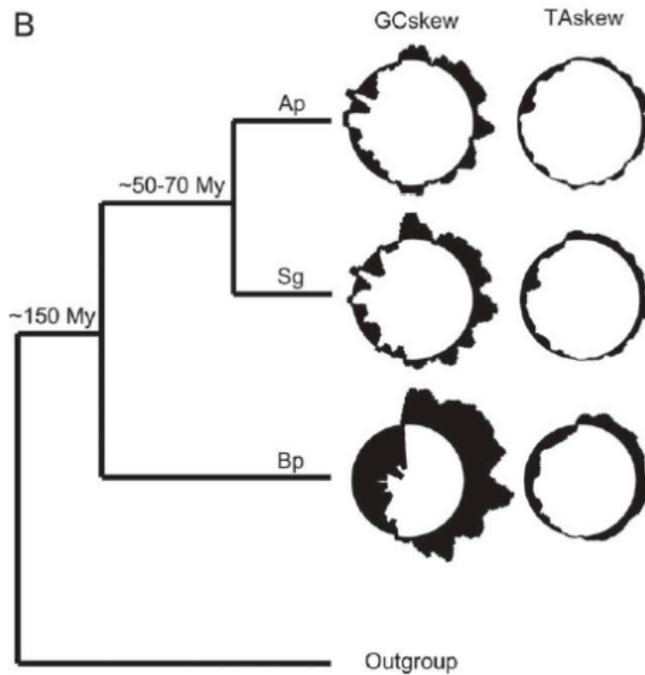
Simple DNA walk *E. coli*

Oriloc



Cytosine deamination theory



Buchnera aphidicola

Chirochores practical

Study this yourself :

- Plot the results of `oriloc()` for *Chlamydia trachomatis* (`library(seqinr)`)
- Have a look at `oriloc()` outputs there :
<http://pbil.univ-lyon1.fr/software/Oriloc/>.

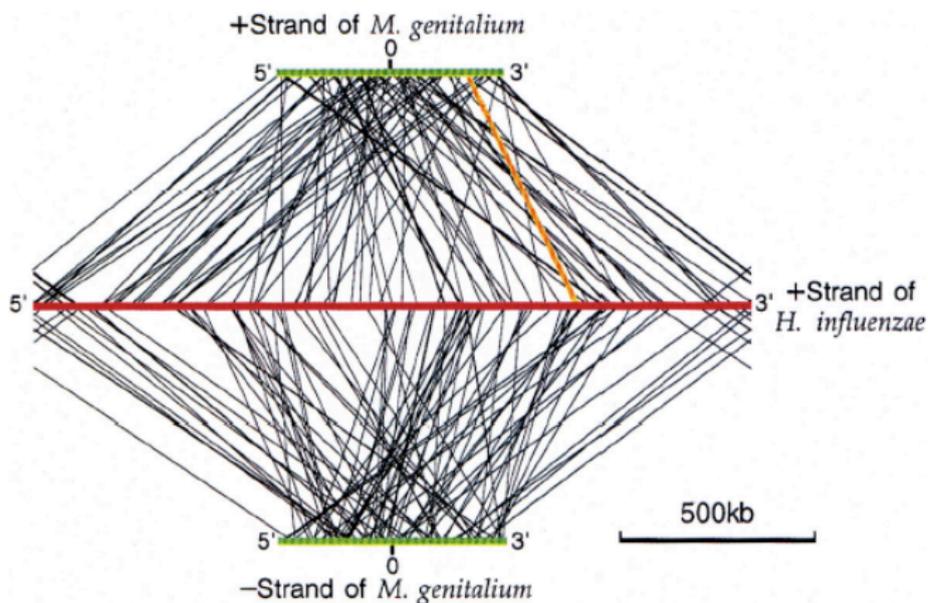
X-rated structure : gene order evolution

- 1 Introduction
- 2 Genome size
- 3 Topology & #
- 4 G+C content
- 5 Replichores
- 6 Gene orientation biases
- 7 Chirochores
- 8 X-rated structure**

Tsuzumi drum

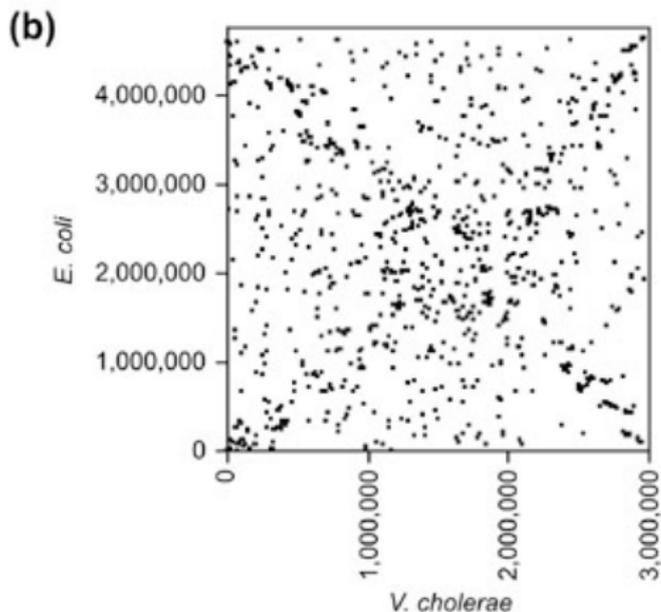


Gene order comparison in bacteria

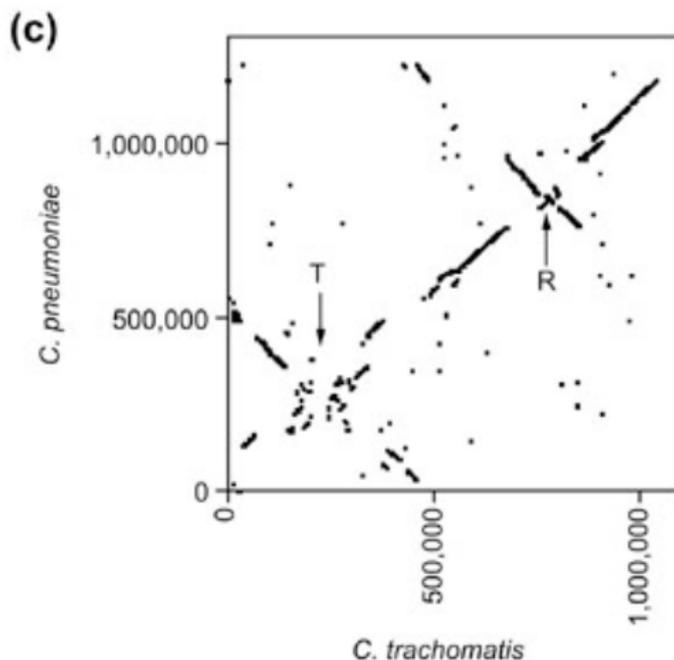


Watanabe *et al.* (1997) *J. Mol. Evol.*, **44** :s57-s64.

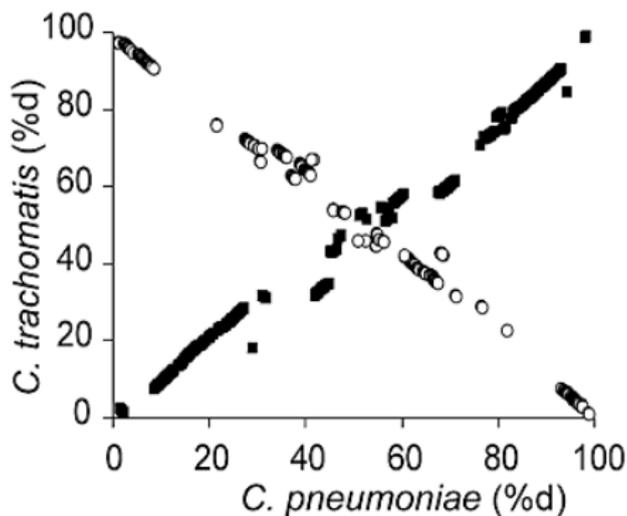
Genomic dot plots (*E. coli* vs. *V. cholerae*)



Eisen *et al.* (2000) *Genome Biology*, **1** :research0011.1-9

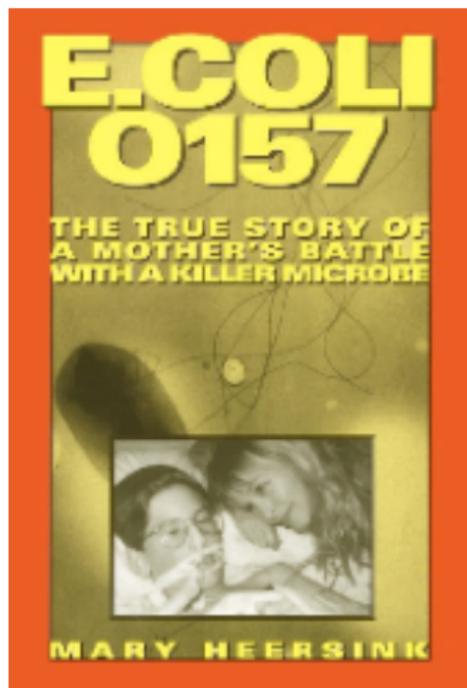
Genomic dot plots (*C. pneumoniae* vs. *C. trachomatis*)

Eisen *et al.* (2000) *Genome Biology*, **1** :research0011.1-9.

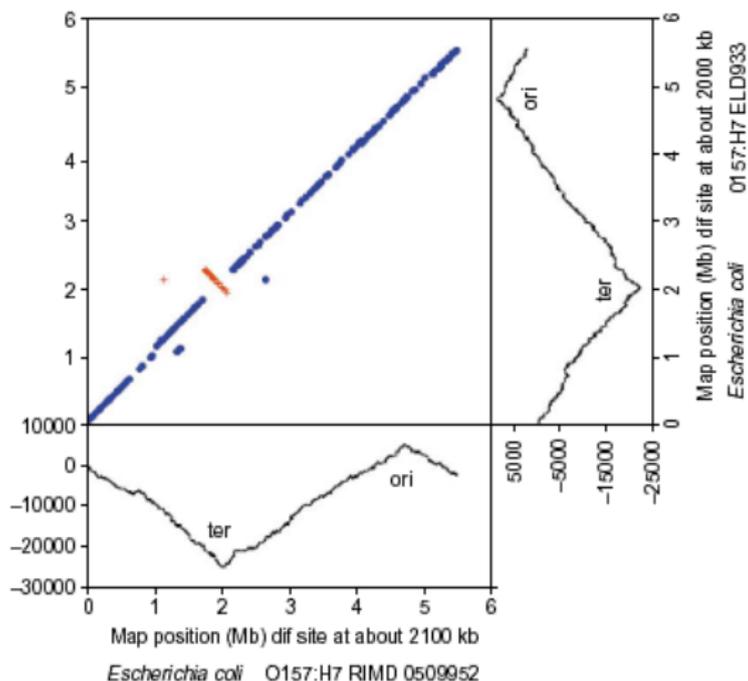
Genomic dot plots (*C. pneumoniae* vs. *C. trachomatis*)

Tillier & Collins (2000) *Nature Genetics*, **26** :195-197.

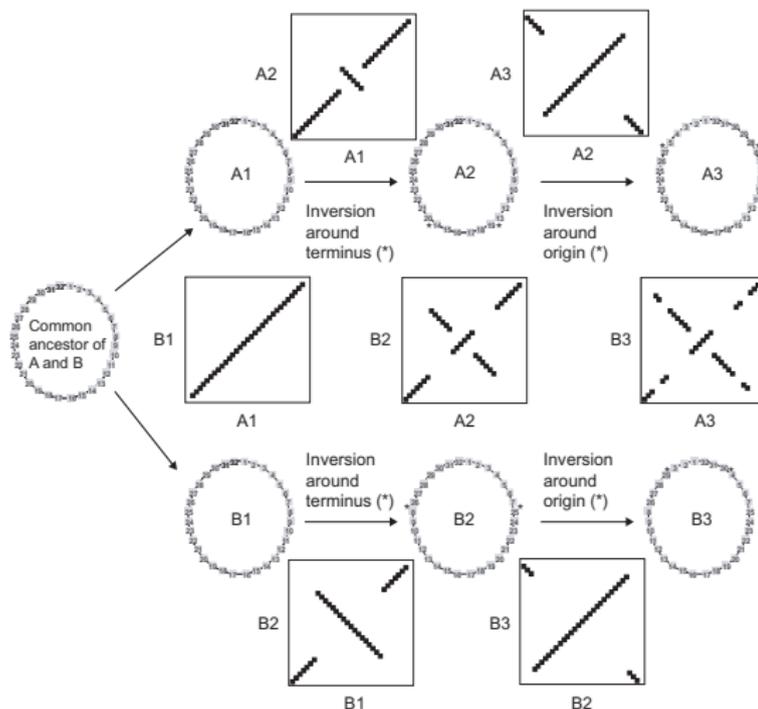
Genomic dot plots (O157 :H7 vs. O157 :H7)



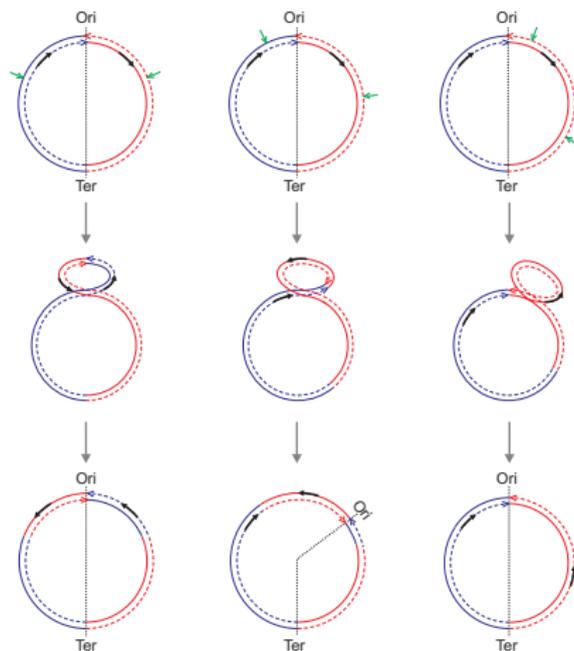
Genomic dot plots (O157 :H7 vs. O157 :H7)



Simulation of symmetric inversions



Three models for inversions



"Bacterial" Genome structures

4BIM - INSA 2006

J. R. Lobry

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18 juin 2006